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(54) Title: OB FUSION PROTEIN COMPOSITIONS AND MET

(57) Abstract

The present invention relates to Fc-OB fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic or chemical fusion protein comprising the Fc immunoglobulin region, derivative or analog fused to the N-terminal portion of the OB protein, derivative or analog.

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OB FUSION PROTEIN COMPOSITIONS AND METHODS

Field of the Invention

The present invention relates to Fc-OB fusion protein compositions and methods for preparation and use thereof

Background

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- Although the molecular basis for obesity is largely unknown, the identification of the "OB gene" and protein encoded ("OB protein" or "leptin") has shed some light on mechanisms the body uses to regulate body fat deposition. See, PCT publication, WO 96/05309
- 15 (12/22/96), Friedman et al.; Zhang et al., Nature 372: 425-432 (1994); see also, the Correction at Nature 374: 479 (1995). The OB protein is active in vivo in both ob/ob mutant mice (mice obese due to a defect in the production of the OB gene product) as well as in normal,
- wild type mice. The biological activity manifests itself in, among other things, weight loss. <u>See</u> generally, Barrinaga, "Obese" Protein Slims Mice, Science <u>269</u>: 475-456 (1995). The OB protein, derivatives and use thereof as modulators for the
- 25 control of weight and adiposity of animals, including mammals and humans, has been disclosed in greater detail in PCT publication WO 96/05309 (12/22/96), hereby incorporated by reference, including figures.
- The other biological effects of OB protein are
 not well characterized. It is known, for instance, that
 in ob/ob mutant mice, administration of OB protein
 results in a decrease in serum insulin levels, and serum
 glucose levels. It is also known that administration of
 OB protein results in a decrease in body fat. This was
- 35 observed in both ob/ob mutant mice, as well as non-obese normal mice. Pelleymounter et al., Science 269: 540-543

(1995); Halaas et al., Science 269: 543-546 (1995). See also, Campfield et al., Science 269: 546-549 (1995) (Peripheral and central administration of microgram doses of OB protein reduced food intake and body weight of ob/ob and diet-induced obese mice but not in db/db obese mice.) In none of these reports have toxicity's been observed, even at the highest doses.

Despite the promise of clinical application of the OB protein, the mode of action of the OB protein in 10 yivo is not clearly elucidated. Information on the OB receptor, shows high affinity binding of the OB protein detected in the rat hypothalamus, which indicates OB receptor location. Stephens et al., Nature 377: 530-532. The db/db mouse displays the identical phenotype as the ob/ob mouse, i.e., extreme obesity and Type II diabetes; this phenotype is thought to be due to a defective OB receptor, particularly since db/db mice fail to respond to OB protein administration. See Stephens et al., supra.

With the advances in recombinant DNA

technologies, the availability of recombinant proteins for therapeutic use has engendered advances in protein formulation and chemical modification. One goal of such modification is protein protection and decreased degradation. Fusion proteins and chemical attachment may effectively block a proteolytic enzyme from physical contact with the protein backbone itself, and thus prevent degradation. Additional advantages include, under certain circumstances, increasing the stability, oriculation time, and the biological activity of the therapeutic protein. A review article describing protein modification and fusion proteins is Francis.

Focus on Growth Factors 3:4-10 (May 1992) (published by Mediscript, Mountview Court, Friern Barnet Lane, London N20, OLD, UK).

One such modification is the use of the Fc

region of immunoglobulins. Antibodies comprise two
functionally independent parts, a variable domain known
as "Fab", which binds antigen, and a constant domain,
known as "Fc" which provides the link to effector
functions such as complement or phagocytic cells. The

Fc portion of an immunoglobulin has a long plasma halflife, whereas the Fab is short-lived. Capon, et al.,

Nature 337: 525-531 (1989). Therapeutic protein products have been constructed using the Fc domain to provide longer half-15 life or to incorporate functions such as Fc receptor binding, protein A binding, complement fixation and placental transfer which all reside in the Fc proteins of immunoglobulins. Id. For example, the Fc region of an IgG1 antibody has been fused to the N-terminal end of 20 CD30-L, a molecule which binds CD30 receptors expressed on Hodgkin's Disease tumor cells, anaplastic lymphoma cells. T-cell leukemia cells and other malignant cell types. See, U.S. Patent No. 5,480,981. IL-10, an antiinflammatory and antirejection agent has been fused to 25 murine Fcy2a in order to increase the cytokine's short circulating half-life. Zheng, X. et al., The Journal of Immunology, 154: 5590-5600 (1995). Studies have also evaluated the use of tumor necrosis factor receptor linked with the Fc protein of human IgG1 to treat 30 patients with septic shock. Fisher, C. et al., N. Engl. J. Med., 334: 1697-1702 (1996); Van Zee, K. et al., The Journal of Immunology, 156: 2221-2230 (1996). Fc has also been fused with CD4 receptor to produce a therapeutic protein for treatment of AIDS. See, Capon

et al., Nature, <u>337</u>:525-531 (1989). In addition, the N-terminus of interleukin 2 has also been fused to the

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PCT/IIS97/23183 WO 98/28427 -4-

Fc portion of IgG1 or IgG3 to overcome the short half life of interleukin 2 and its systemic toxicity. See, Harvill et al., Immunotechnology, 1: 95-105 (1995).

Due to the identification of the OB protein as a promising therapeutic protein, there exists a need to develop OB analog compositions for clinical application in conjunction with or in place of OB protein administration. Such development would include OR analog compositions where protein formulations and 10 chemical modifications achieve decreased protein degradation, increased stability and circulation time. The present invention provides such compositions.

Summary of the Invention

- 15 The present invention relates to Fc-OB fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic fusion protein comprising the Fc region or analogs of immunoglobulins 20 fused to the N-terminal portion of the OB protein or analogs. The Fc-OB fusion protein is capable of dimerizing via the cysteine residues of the Fc region. Unexpectedly, genetic fusion modification with Fc at the N-terminus of the OB protein demonstrates advantages in 25 stability, clearance rate and decreased degradation which are not seen in OB protein or with fusion of Fc to the C-terminus of the OB protein. Surprisingly and importantly, the N-terminus modification provides unexpected protein protection from degradation. 30 increases circulation time and stability, when compared
- to the OB protein or Fc modification to the OB protein C-terminus. Such unexpected advantages from the Fc modification to OB protein would be advantageous to OB protein consumers, in that these changes contribute to lower doses requir d or less frequent dosing. Thus, as
- 35 described below in more detail, the present invention

has a number of aspects relating to the genetic modification of proteins via fusion of the Fc region to the OB protein (or analogs thereof), as well as, specific modifications, preparations and methods of use thereof.

Accordingly, in one aspect, the present invention provides a Fc-OB fusion protein wherein Fc is genetically fused to the N-terminus of the OB protein (or analogs thereof). In addition, the Fc portion may also be linked to the N-terminus of the OB protein (or 10 analogs thereof) via peptide or chemical linkers as known in the art. As noted above and described in more detail below, the Fc-OB fusion protein has unexpected protections from degradation and increased circulation time and stability when compared to the OB protein or 15 C-terminus OB-Fc fusion proteins. Additional aspects of the present invention, therefore, include not only Fc-OB fusion protein compositions, but also DNA sequences encoding such proteins, related vectors and host cells 20 containing such vectors, both useful for producing fusion proteins of the present invention.

In a second aspect, the present invention provides for preparing the Fc-OB fusion protein. Such methods include recombinant DNA techniques for

25 preparation of recombinant proteins. Furthermore, such aspects include methods of fermentation and purification as well.

In another aspect, the present invention provides methods for treating excess weight in an 30 individual or animals, including modulation of and/or fat deposition by the administration of Fc-OB fusion proteins. Due to the Fc-OB fusion protein characteristics, methods are contemplated which reduce the amount and/or frequency of dosage of OB protein by using Fc-OB weight reducing agent.

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In yet another aspect, the present invention provides for therapies for the treatment of co-morbidities associated with excess fat, such as diabetes, dys- or hyperlipidemias, arterial sclerosis, arterial plaque, the reduction or prevention of gall

arterial plaque, the reduction or prevention of gall stones formation, stoke, and also an increase in insulin sensitivity and/or an increase in lean tissue mass.

In another aspect, the present invention also

provides for related pharmaceutical compositions of the 10 Fc-OB proteins, analogs and derivatives thereof, for use in the above therapies.

Brief Description of the Drawings

FIGURE 1 Recombinant murine metOB (double 5 stranded) DNA (SEQ. ID. NOs.: 1 and 2) and amino acid sequence (SEQ. ID. NO. 3).

FIGURE 2 Recombinant human metOB analog (double stranded) DNA (SEQ. ID. NOs.: 4 and 5) and amino acid sequence (SEQ. ID. NO. 6).

FIGURE 3 (A-C) Recombinant human metFc-OB (double stranded) DNA (SEQ. ID. NOs.: 7 and 8) and amino acid sequence (SEQ. ID. NO. 9).

FIGURE 4 (A-C) Recombinant human metFc-OB variant (double stranded) DNA (SEQ. ID. NOs.: 10 and 11) and amino acid sequence (SEQ. ID. NO. 12).

FIGURE 5 (A-C) Recombinant human metFc-OB variant (double stranded) DNA (SEQ. ID. NOs.: 13 and 14) and amino acid sequence (SEQ. ID. NO. 15).

FIGURE 6 (A-C) Recombinant human metFc-OB

variant (double stranded) DNA (SEQ. ID. NOs.: 16 and 17)

and amino acid sequence (SEQ. ID. NO. 18).

Detailed Description

The present invention relates to Fc-OB fusion 5 protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to the genetic or chemical fusion of the Fc region of immunoglobulins to the N-terminal portion of the OB protein. Unexpectedly, fusion of Fc at the N-terminus of the OB protein of demonstrates advantages which are not seen in OB protein or with fusion of Fc at the C-terminus of the OB protein. Surprisingly, the N-terminally modified Fc-OB protein provides unexpected protein protection from degradation, increased circulation time and increased stability. Accordingly, the Fc-OB fusion protein, and analogs or derivatives thereof, as well as, related methods of use and preparation, are described in more detail below.

15 Compositions

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The Fc sequence of the recombinant human Fc-OB sequence set forth in SEQ. ID. NO. 9 (See Figure 3) may be selected from the human immunoglobulin IgG-1 heavy chain, <u>see</u> Ellison, J.W. et al., Nucleic Acids Res. 10: 4071-4079 (1982), or any other Fc sequence known in the art (e.g. other IgG classes including but not limited to IgG-2, IgG-3 and IgG-4, or other immunoglobulins). Variant, analogs or derivatives of the Fc portion may be constructed by, for example, making various substitutions of residues or sequences.

Cysteine residues can be deleted or replaced with other amino acids to prevent formation of disulfide crosslinks of the Fc sequences. In particular amino acid at position 5 of SEQ. ID. NO. 9 is a cysteine residue. The recombinant Fc-OB sequence of SEQ. ID.

NO. 9 is a 378 amino acid Fc-OB protein (not counting the methionine r sidue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 3 is referred to as +1 with the methionine at the -1 position.

One may remove the cysteine residue at position 5 or substitute it with one or more amino acids. An alanine residue may be substituted for the cysteine residue at position 6 giving the variant amino acid sequence of Figure 4 (SEQ. ID. NO. 12). The recombinant Fc-OB protein of Figure 4 is a 378 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 4 is referred to as +1 with the methionine at the -1 position.

Likewise, the cysteine at position 5 of SEO. TO NO 9 could be substituted with a serine or other amino acid residue or deleted. A variant or analog may also be prepared by deletion of amino acids at positions 1. 2. 3. 4 and 5 as with the variant in SEO, ID, NO. 15 (See Figure 5). Substitutions at these positions can also be made and are with in the scope of this invention. The recombinant Fc-OB protein of Figure 5 is a 373 amino acid Fc-OB protein (not counting the 20 methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 5 is referred to as +1 with the methionine at the -1 position.

Modifications may also be made to introduce four amino acid substitutions to ablate the Fc receptor binding site and the complement (Clg) binding site. 25 These variant modifications from SEQ. ID. NO. 15 would include leucine at position 15 substituted with glutamate, glutamate at position 98 substituted with alanine, and lysines at positions 100 and 102

substituted with alanines (see Figure 6 and SEQ. ID. NO. 30 18). The recombinant Fc-OB protein of Figure 6 is a 373 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 6 is referred to as 35 +1 with the methionine at the -1 position.

Likewise, one or more tyrosine residues can be replaced by phenyalanine residues as well. In addition, other variant amino acid insertions, deletions and/or substitutions are also contemplated and are within the scope of the present invention. Furthermore, alterations may be in the form of altered amino acids, such as peptidomimetics or D-amino acids. The Fc protein may be also linked to the OB proteins of the Fc-OB protein by "linker" moieties whether chemical or amino acids of varying lengths. Such chemical linkers are well known in the art. Amino acid linker sequences can include but are not limited to:

(a) ala, ala, ala: (b) ala, ala, ala, ala; 15 (C) ala, ala, ala, ala, ala; (d) glv. glv: (e) glv, glv, glv; gly, gly, gly, gly, gly; (f) (a) gly, gly, gly, gly, gly, gly, gly; 20 (h) gly-pro-gly; (i) gly, gly, pro, gly, gly; and any combination of subparts (a) (i) through (i).

The OB portion of the Fc-OB fusion protein may be selected from the recombinant murine set forth in 25 SEQ. ID. NO. 3 (See Figure 1), or the recombinant human protein as set forth in Zhang et al., Nature, supra, (herein incorporated by reference) or those lacking a glutaminyl residue at position 28. (See Zhang et al, Nature, supra, at page 428.) One may also use the 30 recombinant human OB protein analog as set forth in SEQ. ID. NO. 6 (See Figure 2), which contains: (1) an arginine in place of lysine at position 35; and (2) a leucine in place of isoleucine at position 74. (A 35 shorthand abbreviation for this analog is the recombinant human $R->L^{35}$, $I->L^{74}$). The amino acid

sequences for the recombinant human and recombinant murine proteins or analogs with or without the fused Fc portion at the N-terminus of the OB protein are set forth below with a methionyl residue at the -1 position; however, as with any of the present OB proteins and analogs, the methionyl residue may be absent.

The murine protein is substantially homologous to the human protein, particularly as a mature protein, and, further, particularly at the N-terminus. One may 10 prepare an analog of the recombinant human protein by altering (such as substituting amino acid residues), in the recombinant human sequence, the amino acids which diverge from the murine sequence. Because the recombinant human protein has biological activity in 15 mice, such an analog would likely be active in humans. For example, using a human protein having a lysine at residue 35 and an isoleucine at residue 74 according to the numbering of SEQ. ID. NO. 6, wherein the first amino acid is valine, and the amino acid at position 146 is cysteine, one may substitute with another amino acid one 20 or more of the amino acids at positions 32, 35, 50, 64, 68, 71, 74, 77, 89, 97, 100, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145. One may select the amino acid at the corresponding position of the murine protein, (SEQ. ID. NO. 3), or another amino acid. 25

One may further prepare "consensus" molecules based on the rat OB protein sequence. Murakami et al., Biochem. Biophys. Res. Comm. 202: 944-952 (1995) herein incorporated by reference. Rat OB protein differs from 100 protein at the following positions (using the numbering of SEQ. ID. NO. 6): 4, 12, 33, 15, 50, 68, 71, 74, 77, 78, 89, 97, 100, 101, 102, 105, 106, 107, 108, 111, 118, 136, 138 and 145. One may substitute with another amino acid one or more of the amino acids at

35 these divergent positions. The positions in bold print are those in which the murine OB protein as well as the

rat OB protein are divergent from the human OB protein, and thus, are particularly suitable for alteration. At one or more of a positions, one may substitute an amino acid from the corresponding rat OB protein, or another amino acid.

The positions from both rat and murine OB protein which diverge from the mature human OB protein are: 4, 32, 33, 35, 50, 64, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145. An OB protein according to SEQ. ID. NO. 6 having one or more of the above amino acids replaced with another amino acid, such as the amino acid found in the corresponding rat or murine sequence, may also be effective.

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15 In addition, the amino acids found in rhesus monkey OB protein which diverge from the mature human OB protein are (with identities noted in parentheses in one letter amino acid abbreviation): 8 (S), 35 (R), 48(V), 53(Q), 60(I), 66(I), 67(N), 68((L), 89(L), 100(L), 20 108(E), 112 (D), and 118 (L). Since the recombinant human OB protein is active in cynomolgus monkeys, a human OB protein according to SEO, ID, NO. 6 (with lysine at position 35 and isoleucine at position 74) having one or more of the rhesus monkey divergent amino 25 acids replaced with another amino acid, such as the amino acids in parentheses, may be effective. It should be noted that certain rhesus divergent amino acids are also those found in the above murine species (positions 35, 68, 89, 100 and 112). Thus, one may prepare a

- murine/rhesus/human consensus molecule having (using the numbering of SEQ. ID. NO. 6 having a lysine at position 35 and an isoleucine at position 74) having one or more of the amino acids at positions replaced by another amino acid: 4, 8, 32, 33, 35, 48, 50, 53, 60, 64, 66,
- 35 67, <u>68</u>, 71, 74, 77, 78, <u>89</u>, 97, <u>100</u>, 102, 105, 106, 107, 108, 111, <u>112</u>, 118, 136, 138, 142, and 145.

Other analogs may be prepared by deleting a part of the protein amino acid sequence. For example, the mature protein lacks a leader sequence (-22 to -1). One may prepare the following truncated forms of human OB protein molecules (using the numbering of SEQ. ID. NO. 6):

- (a) amino acids 98-146
- (b) amino acids 1-32
- (c) amino acids 40-116
- (d) amino acids 1-99 and (connected to)

112-146

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- (e) amino acids 1-99 and (connected to)
 112-146 having one or more of amino acids 100-111 placed
 between amino acids 99 and 112.
- In addition, the truncated forms may also have altered one or more of the amino acids which are divergent (in the rat, murine, or rhesus OB protein) from human OB protein. Furthermore, any alterations may be in the form of altered amino acids, such as peptidomimetics or D-amino acids.

Therefore, the present invention encompasses a Fc-OB fusion protein wherein the OB protein is selected from:

- (a) the amino acid sequence 1-146 as set
- 25 forth in SEQ. ID. NO. 3 (below) or SEQ. ID. NO. 6;
 - (b) the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 6 having a lysine residue at position 35 and an isoleucine residue at position 74; (c) the amino acid sequence of subpart (b)
- 30 having a different amino acid substituted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 6 and retaining the same numbering even in the absence of a glutaminyl residue at position 28): 4, 32, 33, 35, 50, 64, 68, 71, 74, 77,
- 35 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145;

WO 98/28427

-13-

PCT/US97/23183

(d) the amino acid sequence of subparts (a). (b) or (c) optionally lacking a glutaminyl residue at position 28: (e) the amino acid sequence of subparts (a), (b), (c), or (d) having a methionvl residue at the N-terminus: (f) a truncated OB protein analog selected from among: (using the numbering of SEO, ID, NO, 6): (i) amino acids 98-146 (ii) amino acids 1-32 10 (iii) amino acids 40-116 (iv) amino acids 1-99 and 112-146 (v) amino acids 1-99 and 112-146 having one or more of amino acids 100-111 placed between 15 amino acids 99 and 112; and, (vi) the truncated OB analog of subpart (i) having one or more of amino acids 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145 substituted with another amino acid: 20 (vii) the truncated analog of subpart (ii) having one or more of amino acids 4, 8 and 32 substituted with another amino acid: (viii) the truncated analog of subpart (iii) having one or more of amino acids 50, 53, 60, 25 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111 and 112 replaced with another amino acid: (vix) the truncated analog of subpart (iv) having one or more of amino acids 4, 8, 32, 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 30 78. 89. 97, 112, 118, 136, 138, 142, and 145 replaced with another amino acid; and

(x) the truncated analog of subpart (v)

having one or more of amino acids 4, 32, 33, 35,

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- 50, 64, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145 replaced with another amino acid;
- (xi) the truncated analog of any of subparts (i)-(x) having an N-terminal methionyl residue; and
- (g) the OB protein or analog derivative of any of subparts (a) through (f) comprised of a chemical moiety connected to the protein moiety;
- (h) a derivative of subpart (g) wherein said chemical moiety is a water soluble polymer moiety;
- (i) a derivative of subpart (h) wherein said water soluble polymer moiety is polyethylene glycol;
- (j) a derivative of subpart (h) wherein said 15 water soluble polymer moiety is a polyaminoacid moiety;
 - (k) a derivative of subpart (h) through (j)wherein said moiety is attached at solely the N-terminus of said protein moiety; and
- (1) an OB protein, analog or derivative of 20 any of subparts (a) through (k) in a pharmaceutically acceptable carrier.

Derivatives

- The present Fc-OB fusion proteins (herein the

 25 term "protein" is used to include "peptide," Fc, OB or
 analogs, such as those recited infra, unless otherwise
 indicated) are derivatized by the attachment of one or
 more chemical moieties to the Fc-OB fusion protein
 moiety. These chemically modified derivatives may be

 30 further formulated for intraarterial, intraperitoneal,
 intramegular subgutaneous, intraperous, oral, pasal.
- intramuscular subcutaneous, intravenous, oral, nasal, pulmonary, topical or other routes of administration as discussed below. Chemical modification of biologically active proteins has been found to provide additional
- 35 advantages under certain circumstances, such as increasing the stability and circulation time of the

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PCT/US97/23183

therapeutic protein and decreasing immunogenicity. <u>Sec</u>, U.S. Patent No. 4,179,337, Davis et al., issued December 18, 1979. For a review, <u>see</u> Abuchowski et al., in Enzymes as Drugs. (J. S. Holcerberg and J. Roberts, eds. pp. 367-383 (1981)); Francis et al., supra.

The chemical moieties suitable for such derivatization may be selected from among various water soluble polymers. The polymer selected should be water soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a physiological environment. Preferably, for therapeutic use of the end-product preparation, the polymer will be pharmaceutically acceptable. One skilled in the art will be able to select the desired polymer based on such considerations as whether the polymer/protein conjugate will be used therapeutically, and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. For the present proteins and peptides, the effectiveness of the derivatization may be ascertained by administering the derivative, in the desired form (i.e., by osmotic pump, or, more preferably, by injection or infusion, or, further formulated for oral, pulmonary or nasal delivery, for example), and observing biological effects as described

herein.

The water soluble polymer may be selected from the group consisting of, for example, polyethylene glycol, copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol,

- 30 polyvinyl pyrolidone, poly-1, 3-dioxolane, poly-1,3,6-trioxane, ethylene/maleic anhydride copolymer, polyaminoacids (either homopolymers or random copolymers), and dextran or poly(n-vinyl pyrolidone)polyethylene glycol, propylene glycol
- 35 homopolymers, polypropylene oxide/ethylene oxide co-polymers, polypropylene oxide/ethylene oxide

alcohol. Polyethylene glycol propionaldenhyde may have advantages in manufacturing due to its stability in water. Also, succinate and styrene may also be used.

The OB or Fc proteins used to formulate the

5 Fc-OB fusion protein, may be prepared by attaching polyaminoacids or branch point amino acids to the Fc or OB protein (or analogs) moiety. For example, the polyaminoacid may be an additional carrier protein which, like the Fc fused to the OB protein or OB analog 10 of the present invention, serves to also increase the circulation half life of the protein in addition to the advantages achieved via the Fc-OB fusion protein above. For the present therapeutic or cosmetic purpose of the present invention, such polyaminoacids should be those 15 which have or do not create neutralizing antigenic response, or other adverse responses. Such polyaminoacids may be selected from the group consisting of serum album (such as human serum albumin), an additional antibody or portion thereof (e.g. the Fc region), or other polyaminoacids, e.g. lysines. As

20 region), or other polyaminoacids, e.g. lysines. As indicated below, the location of attachment of the polyaminoacid may be at the N-terminus of the FC-OB protein moiety, or C-terminus, or other places in between, and also may be connected by a chemical

25 "linker" moiety to the FC-OB protein.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 2 kDa and about 100 kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic profile (e.g., the duration of sustained rel ase desired, the eff cts, if any on biological activity, the ease in handling, the degree or

lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or analog).

The number of polymer molecules so attached may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-derivatize, or may provide for a di-, tri-, tetraor some combination of derivatization, with the same or different chemical moieties (e.g., polymers, such as different weights of polyethylene glycols). proportion of polymer molecules to protein (or peptide) molecules will vary, as will their concentrations in the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization 15 (e.g., mono, di-, tri-, etc.), the molecular weight of the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

The chemical mojeties should be attached to

the protein with consideration of effects on functional

or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art. E.g., EP 0 401 384 herein incorporated by reference (coupling PEG to G-CSF), see also Malik et al., Exp. Hematol. 20: 1028-1035 (1992) (reporting pegylation of 25 GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those 30 to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid residue. Those having a free carboxyl group may include aspartic acid residues, glutamic acid 35 residues, and the C-terminal amino acid residue.

Sulfhydryl groups may also be used as a reactive group

-18-

for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues important for receptor binding should be avoided if receptor binding is desired.

One may specifically desire N-terminally chemically modified Fc-OB fusion protein. Using polyethylene glycol as an illustration of the present compositions, one may select from a variety of 10 polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be 15 performed, and the method of obtaining the selected N-terminally pegylated protein. The method of obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated 20 protein molecules. Selective N-terminal chemical modification may be accomplished by reductive alkylation

types of primary amino groups (lysine versus the

N-terminal) available for derivatization in a particular
protein. Under the appropriate reaction conditions,
substantially selective derivatization of the protein at
the N-terminus with a carbonyl group containing polymer
is achieved. For example, one may selectively

which exploits differential reactivity of different

30 N-terminally pegylate the protein by performing the reaction at a pH which allows one to take advantage of the pK_B differences between the ε-amino group of the lysine r sidues and that of the α-amino group of the N-terminal r sidue of the protein. By such selective derivatization, attachment of a water soluble polymer to a protein is controlled: the conjugation with the

-19-

polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water 5 soluble polymer may be of the type described above, and should have a single reactive aldehyde for coupling to the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

An N-terminally monopegylated derivative is
preferred for ease in production of a therapeutic.
N-terminal pegylation ensures a homogenous product as characterization of the product is simplified relative to di-, tri- or other multi-pegylated products. The use of the above reductive alkylation process for preparation of an N-terminal product is preferred for ease in commercial manufacturing.

Complexes

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The Fc-OB fusion protein, analog or derivative thereof may be administered complexed to a binding 20 composition. Such binding composition may have the effect of prolonging the circulation time even further than that achieved with the Fc-OB fusion protein, analog or derivative. Such composition may be a protein 25 (or synonymously, peptide). An example of a binding protein is OB protein receptor or portion thereof, such as a soluble portion thereof. Other binding proteins may be ascertained by examining OB protein or Fc-OB protein in serum, or by empirically screening for the presence of binding. Binding proteins used will typically not interfere with the ability of OB protein, Fc-OB fusion proteins, or analogs or derivatives thereof, to bind to endogenous OB protein receptor and/or effect signal transduction.

PCT/US97/23183 WO 98/28427

-20-

Pharmaceutical Compositions

The present invention also provides methods of using pharmaceutical compositions of the Fc-OB fusion proteins and derivatives. Such pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal, transdermal or other forms of administration. In general, comprehended by the invention are pharmaceutical compositions comprising effective amounts of protein or derivative products of 10 the invention together with pharmaceutically acceptable diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 15 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into 20 particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the 25 physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. See, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference. The compositions may be prepared in liquid 30 form, or may be in dried powder, such as lyophilized form. Implantable sustained release formulations are also contemplated, as are transdermal formulations. Contemplated for use herein are oral solid

35 dosage forms, which are described generally in Remington's Pharmaceutical Sciences, 18th Ed. 1990 (Mack

Publishing Co. Easton PA 18042) at Chapter 89, which is herein incorporated by reference. Solid dosage forms include tablets, capsules, pills, troches or lozenges, cachets or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673). Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (e.g., U.S. Patent No. 5.013.556). A description of possible solid dosage 10 forms for the therapeutic is given by Marshall, K. In: Modern Pharmaceutics Edited by G. S. Banker and C. T. Rhodes Chapter 10, 1979, herein incorporated by reference. In general, the formulation will include the 15 Fc-OB fusion protein (or analog or derivative), and inert ingredients which allow for protection against the stomach environment, and release of the biologically

Also specifically contemplated are oral dosage

20 forms of the above derivatized proteins. Fc-OB fusion protein may be chemically modified so that oral delivery of the derivative is efficacious. Generally, the chemical modification contemplated is the attachment of at least one moiety to the protein (or peptide) molecule 25 itself, where said moiety permits (a) inhibition of proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also desired is the increase in overall stability of the protein and increase in circulation time in the body. Examples of such moieties 30 include: Polyethylene glycol, copolymers of ethylene glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone and polyproline. Abuchowski and Davis, Soluble Polymer-Enzyme Adducts. In: "Enzymes as Drugs". 35 Hocenberg and Roberts, eds., Wiley-Interscience, New York, NY, (1981), pp. 367-383; Newmark, et al., J. Appl.

active material in the intestine.

WO 98/28427

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Biochem. 4: 185-189 (1982). Other polymers that could be used are poly-1,3-dioxolane and poly-1,3,6-tioxocane. Preferred for pharmaceutical usage, as indicated above. are polyethylene glycol moieties.

PCT/I/S07/23183

For the Fc-OB fusion protein, analog or derivative, the location of release may be the stomach, the small intestine (e.g., the duodenum, jejunum, or ileum), or the large intestine. One skilled in the art has available formulations which will not dissolve in the stomach, yet will release the material in the duodenum or elsewhere in the intestine. Preferably, the release will avoid the deleterious effects of the stomach environment, either by protection of the Fc-OB fusion protein, analog or derivative, or by release of the biologically active material beyond the stomach environment, such as in the intestine.

To ensure full gastric resistance a coating impermeable to at least pH 5.0 is essential. Examples of the more common inert ingredients that are used as 20 enteric coatings are cellulose acetate trimellitate (CAT), hydroxypropylmethylcellulose phthalate (HPMCP), HPMCP 50, HPMCP 55, polyvinyl acetate phthalate (PVAP). Eudragit L30D, Aquateric, cellulose acetate phthalate (CAP), Eudragit L, Eudragit S, and Shellac. These 25 coatings may be used as mixed films.

A coating or mixture of coatings can also be used on tablets, which are not intended for protection against the stomach. This can include sugar coatings. or coatings which make the tablet easier to swallow. Capsules may consist of a hard shell (such as gelatin)

- 30 for delivery of dry therapeutic i.e. powder; for liquid forms, a soft gelatin shell may be used. The shell material of cach ts could be thick starch or other edible paper. For pills, lozenges, molded tablets or
- 35 tablet triturates, moist massing techniques can be used.

-23-

The therapeutic can be included in the formulation as fine multiparticulates in the form of granules or pellets of particle size about 1 mm. The formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by compression.

Colorants and flavoring agents may all be included. For example, the protein (or derivative) may be formulated (such as by liposome or microsphere encapsulation) and then further contained within an edible product, such as a refrigerated beverage containing colorants and flavoring agents.

One may dilute or increase the volume of the 15 therapeutic with an inert material. These diluents could include carbohydrates, especially mannitol. α-lactose, anhydrous lactose, cellulose, sucrose, modified dextrans and starch. Certain inorganic salts may be also be used as fillers including calcium 20 triphosphate, magnesium carbonate and sodium chloride. Some commercially available diluents are Fast-Flo.

Emdex, STA-Rx 1500, Emcompress and Avicell. Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. 25 Materials used as disintegrates include but are not limited to starch including the commercial disintegrant based on starch, Explotab. Sodium starch glycolate. Amberlite, sodium carboxymethylcellulose, ultramylopectin, sodium alginate, gelatin, orange peel, acid carboxymethyl cellulose, natural sponge and 30 bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth. Alginic acid and its sodium

salt are also useful as disintegrants.

Binders may be used to hold the therapeutic agent together to form a hard tablet and include materials from natural products such as acacia, tragacanth, starch and gelatin. Others include methyl cellulose (MC), ethyl cellulose (EC) and carboxymethyl cellulose (CMC). Polyvinyl pyrrolidone (PVP) and hydroxypropylmethyl cellulose (HPMC) could both be used in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the
formulation of the therapeutic to prevent sticking
during the formulation process. Lubricants may be used
as a layer between the therapeutic and the die wall, and
these can include but are not limited to; stearic acid
including its magnesium and calcium salts,
polytetrafluoroethylene (PTFE), liquid paraffin,
vegetable oils and waxes. Soluble lubricants may also
be used such as sodium lauryl sulfate, magnesium lauryl
sulfate, polyethylene glycol of various molecular
weights, Carbowax 4000 and 6000.

Glidants that might improve the flow properties of the drug during formulation and to aid rearrangement during compression might be added. The glidants may include starch, talc, pyrogenic silica and hydrated silicoaluminate.

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To aid dissolution of the therapeutic into the aqueous environment a surfactant might be added as a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or benzethomium chloride. The list of potential nonionic detergents that could be included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate, polyorbate 40, 60, 65 and 80, sucrose fatty acid ester,

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methyl cellulose and carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

Additives which potentially enhance uptake of the protein (or derivative) are for instance the fatty acids oleic acid, linoleic acid and linolenic acid.

Controlled release formulation may be desirable. The drug could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms e.g., gums. Slowly degenerating matrices may also be incorporated into the formulation, e.g., alginates, polysaccahrides. Another form of a controlled release of this therapeutic is by a method based on the Oros therapeutic system (Alza Corp.), i.e., the drug is enclosed in a semipermeable membrane which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which could be applied in a coating pan. The therapeutic agent could also be given in a film coated tablet and the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium carboxy-methyl cellulose, providene and the polyethylene glycols. The second group consists of the enteric 30 materials that are commonly esters of phthalic acid.

A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

Also contemplated herein is pulmonary delivery of the present protein (or derivatives thereof). The protein (or derivative) is delivered to the lungs of a mammal while inhaling and traverses across the lung epithelial lining to the blood stream. (Other reports of this include Adjei et al., Pharmaceutical Research 7: 565-569 (1990); Adjei et al., International Journal of Pharmaceutics 63: 135-144 (1990) (leuprolide acetate): Braquet et al., Journal of Cardiovascular 10 Pharmacology 13 (suppl. 5): s.143-146 (1989) (endothelin-1); Hubbard et al., Annals of Internal Medicine 3: 206-212 (1989) (al-antitrypsin); Smith et al., J. Clin. Invest. 84: 1145-1146 (1989) (α-1-proteinase); Oswein et al., *Aerosolization of Proteins". Proceedings of Symposium on Respiratory 15 Drug Delivery II, Keystone, Colorado, March, 1990 (recombinant human growth hormone); Debs et al., The Journal of Immunology 140: 3482-3488 (1988) (interferon-y and tumor necrosis factor a) and Platz et al., U.S.

factor). Contemplated for use in the practice of this invention are a wide range of mechanical devices designed for pulmonary delivery of therapeutic products. including but not limited to nebulizers, metered dose

Patent No. 5,284,656 (granulocyte colony stimulating

25 inhalers, and powder inhalers, all of which are familiar to those skilled in the art.

Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured by Glaxo Inc., Research Triangle Park, 35 North Carolina; and the Spinhaler powder inhaler.

manufactured by Fisons Corp., Bedford, Massachusetts.

-27-

All such devices require the use of formulations suitable for the dispensing of protein (or analog or derivative). Typically, each formulation is specific to the type of device employed and may involve the use of an appropriate propellant material, in addition to diluents, adjuvants and/or carriers useful in therapy.

The protein (or derivative) should most advantageously be prepared in particulate form with an average particle size of less than 10 mm (or microns). most preferably 0.5 to 5 µm, for most effective delivery to the distal lung.

Carriers include carbohydrates such as trehalose, mannitol, xylitol, sucrose, lactose, and 15 sorbitol. Other ingredients for use in formulations may include DPPC, DOPE, DSPC and DOPC, Natural or synthetic surfactants may be used. Polyethylene glycol may be used (even apart from its use in derivatizing the protein or analog). Dextrans, such as cyclodextran, may be used. Bile salts and other 20

related enhancers may be used. Cellulose and cellulose derivatives may be used. Amino acids may be used, such as use in a buffer formulation.

Also, the use of liposomes, microcapsules or 25 microspheres, inclusion complexes, or other types of carriers is contemplated.

Formulations suitable for use with a nebulizer, either jet or ultrasonic, will typically comprise Fc-OB protein, analogs or derivatives thereof. 30 dissolved in water at a concentration of about 0.1 to 25 mg of biologically active protein per mL of solution. The formulation may also include a buffer and a simple sugar (e.g., for protein stabilization and regulation of osmotic pressure). The nebulizer formulation may also 35 contain a surfactant, to reduce or prevent surface

induced aggregation of the protein caused by atomization of the solution in forming the aerosol.

Formulations for use with a metered-dose inhaler device will generally comprise a finely divided powder containing the protein (or derivative) suspended in a propellant with the aid of a surfactant. The propellant may be any conventional material employed for this purpose, such as a chlorofluorocarbon, a hydrochlorofluorocarbon, a hydrofluorocarbon, or a 10 hydrocarbon, including trichlorofluoromethane, dichlorodifluoromethane, dichlorotetrafluoroethanol, and 1,1,1,2-tetrafluoroethane, or combinations thereof. Suitable surfactants include sorbitan trioleate and soya lecithin. Oleic acid may also be useful as a 15 surfactant.

Formulations for dispensing from a powder inhaler device will comprise a finely divided dry powder containing protein (or derivative) and may also include a bulking agent, such as lactose, sorbitol, sucrose, mannitol, trehalose, or xylitol in amounts which facilitate dispersal of the powder from the device. e.g., 50 to 90% by weight of the formulation.

Nasal delivery of the protein (or analog or derivative) is also contemplated. Nasal delivery allows 25 the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for masal delivery include those with dextran or cyclodextran. Delivery via transport 30 across other mucus membranes is also contemplated.

Dosage

One skilled in the art will be able to ascertain effective dosages by administration and 35 observing the desired therapeutic effect. Due to the N-terminus modification of the OB protein, the present invention provides unexpected protein protection from degradation, and increases circulation time and stability, when compared to OB protein or C-terminus modification of the OB protein. One skilled in the art. therefore, will be able to ascertain from these changes that an effective dosage may require lower doses or less frequent dosing.

Preferably, the formulation of the molecule will be such that between about .10 µg/kg/day and 10 mg/kg/day will yield the desired therapeutic effect. 10 The effective dosages may be determined using diagnostic tools over time. For example, a diagnostic for measuring the amount of OB protein or Fc-OB fusion protein in the blood (or plasma or serum) may first be 15 used to determine endogenous levels of protein. Such diagnostic tools may be in the form of an antibody assay, such as an antibody sandwich assay. The amount of endogenous OB protein is quantified initially, and a baseline is determined. The therapeutic dosages are determined as the quantification of endogenous and 20 exogenous OB protein or Fc-OB fusion protein (that is, protein, analog or derivative found within the body. either self-produced or administered) is continued over the course of therapy. The dosages may therefore vary over the course of therapy, with a relatively high 25 dosage being used initially, until therapeutic benefit is seen, and lower dosages used to maintain the therapeutic benefits.

Ideally, in situations where solely reduction in blood lipid levels is desired, where maintenance of 30 reduction of blood lipid levels is desired, or an increase in lean body mass is desired, the dosage will be insufficient to result in weight loss. Thus, during an initial course of therapy of an obese person, dosages 35 may be administered whereby weight loss and concomitant blood lipid level lowering or concomitant fat tissue

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decrease/lean mass increase is achieved. Once sufficient weight loss is achieved, a dosage sufficient to prevent re-gaining weight, yet sufficient to maintain desired blood lipid levels or lean mass increase (or, prevention of lean mass depletion) may be administered. These dosages can be determined empirically, as the effects of OB or Fc-OB protein are reversible, (e.g., Campfield et al., Science 269: 546-549 (1995) at 547). Thus, if a dosage resulting in weight loss is observed when weight 10 loss is not desired, one would administer a lower dose in order to achieve the desired blood lipid levels or increase in lean tissue mass, yet maintain the desired weight.

For increasing an individual's sensitivity to insulin, similar dosage considerations may be taken into 15 account. Lean mass increase without weight loss may be achieved sufficient to decrease the amount of insulin (or, potentially, amylin, thiazolidinediones, or other potential diabetes treating drugs) an individual would 20 be administered for the treatment of diabetes.

For increasing overall strength, there may be similar dosage considerations. Lean mass increase with concomitant increase in overall strength may be achieved with doses insufficient to result in weight loss. Other benefits, such as an increase in red blood cells (and oxygenation in the blood) and a decrease in bone resorption or osteoporosis may also be achieved in the absence of weight loss.

Combinations

The present methods may be used in conjunction with other medicaments, such as those useful for the treatment of diabetes (e.g., insulin, possibly, thiazolidinediones, amylin, or antagonists thereof), cholesterol and blood pressure lowering medicaments (such as those which reduce blood lipid levels or other

cardiovascular medicaments), and activity increasing medicaments (e.g., amphetamines). Appetite suppressants may also be used (such as those affecting the levels of serotonin or neuropeptide Y). Such administration may be simultaneous or may be in <u>seriatim</u>.

In addition, the present methods may be used in conjunction with surgical procedures, such as cosmetic surgeries designed to alter the overall appearance of a body (e.g., liposuction or laser surgeries designed to reduce body mass). The health 10 benefits of cardiac surgeries, such as bypass surgeries or other surgeries designed to relieve a deleterious condition caused by blockage of blood vessels by fatty deposits, such as arterial plaque, may be increased with 15 concomitant use of the present compositions and methods. Methods to eliminate gall stones, such as ultrasonic or laser methods, may also be used either prior to, during or after a course of the present therapeutic methods. Furthermore, the present methods may be used as an adjunct to surgeries or therapies for broken bones. damaged muscle, or other therapies which would be

improved by an increase in lean tissue mass.

The following examples are offered to more fully illustrate the invention, but are not to be construed as limiting the scope thereof.

EXAMPLE 1: Use of Murine FC-OB Protein Via Subcutaneous Injection

This example demonstrates that injection

subcutaneously of murine Fc-OB protein results in weight loss in normal mice. Normal (non-obese) CD1 mice were administered murine Fc-OB protein via subcutaneous injections over a 22 day time period. A dosage of 10 mg protein/kg body weight/day resulted in a 14% (+/- 1.1%)

loss from baseline weight by the 22nd day of injections. A dosage of PBS resulted in a 3.9% (+/- 3.3%) loss from

baseline weight by the 22nd day of injections. The weight loss with the use of 10 mg protein/kg body weight/day of Fc-OB protein in obese CD1 mice resulted in a 10% (+/- 4.3%) loss from baseline weight and a dosage of PBS resulted in a 8.7% (+/- 1.3%) loss from baseline weight, both by the 22nd day of injections Presented below are the percent (%) differences from baseline weight in CD1 mice (8 weeks

differences from baseline weight in CD1 mice (8 weeks old):

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Table 1
Weight Loss Upon Subcutaneous Injection

Time	Vehicle (PBS)	Lean/Recombinant	Obese/Recombinant
(days)		Fc-OB Fusion	Fc-OB Fusion
		Protein	Protein
1-2	44 +/1.1	-3.6 +/41	1.03 +/- 1.36
3-4	-1.07 +/13	-6.8 +/- 1.5	-2.7 +/- 1.1
5-6	13 +/- 1.1	-9.5 +/- 1.2	-4.9 +/95
7-8	92 +/29	-12.5 +/- 1.6	-7.7 +/- 2.9
9-10	1.6 +/- 1.3	-12.6 +/- 1.9	-8.2 +/- 2.9
11-12	-1.98 +/- 1	-13.6 +/- 1.96	-8.6 +/- 2.9
13-14	-5.2 +/- 1.3	-14.6 +/-1.7	-10.1+/- 3.6
15-16	-8.6 +/- 0.1	-14.5 +/- 2	-9.4 +/- 2.2
17-18	-8.5 +/64	-16.1 +/- 1.8	-9.6 +/- 2.99
19-20	-4.1 +/99	-16 +/- 1.5	-10.4 +/- 3.3
21-22	-3.9 +/- 3.3	-14.1 +/- 1.1	-10 +/- 4.3

As can be seen, at the end of a 22 day subcutaneous regime, animals receiving the FC-OB protein lost over 14.1% of their body weight in lean and 10% of body weight in obese, as compared to animals only receiving the PBS vehicle and as compared to baseline.

Surprisingly, animals receiving Fc-OB protein up to 22 days continued to loose weight up until 28

days, 4 days after the last injection. Normal (nonobese) CDI mice administered 10 mg protein/kg body
weight/day of murine Fc-OB protein via subcutaneous
injections stopped at day 22 resulted in a 21% loss from
5 baseline weight at day 28 as compared to 14% loss at day
22. Likewise, obese CDI mice administered 10 mg
protein/kg body weight/day of murine Fc-OB protein
stopped at day 22 resulted in a 13% loss from baseline
weight at day 28 compared to 10% loss at day 22. At day
10 34 weight loss was maintained at 10% loss in obese mice
where lean mice recovered to 5% loss. Controls in each
system from day 22 through day 34 averaged from 4% in
obese mice and 7% dain in lean mice.

15 EXAMPLE 2: Use of Human FC-OB Protein Via Subcutaneous Injection in C57 Mice

This example demonstrates that injection subcutaneously of human Fc-OB protein results in weight loss in normal mice. Normal (non-obese) C57 mice were administered human Fc-OB protein via subcutaneous injections over a 7 day time period. A dosage of 10 mg protein/kg body weight/day resulted in a 12% (+/- 1.3%) loss from baseline weight by the 7th day of injections. A dosage of 1 mg protein/kg body weight/day resulted in a 8.9% (+/- 1.5%) loss from baseline weight by the 7th day of injections. The weight loss with the use of 10 mg protein/kg body weight/day of human OB protein in obese C57 mice resulted in a 1.1% (+/- .99%) loss from baseline weight and a dosage of 1 mg protein/kg body

Results

Presented below are the percent (%)

35 differences from baseline weight in C57 mice (8 weeks old):

30 weight/day resulted in a 2.5% (+/- 1.1%) loss from baseline weight, both by the 7th day of injections.

Table 2
Weight Loss Upon Subcutaneous Injection

Time	Vehicle (PBS)	Recombinant	Recombinant OB
(days)		Fc-OB Fusion	Protein
		Protein	
1-2	.258 +/- 1.3	-6.4 +/- 1.6	-2.1 +/91
3-4	2.2 +/- 1.1	-12.1 +/- 1.5	78 +/36
5-6	4.5 +/- 2	-11.5 +/- 1.5	-1.7 +/6
7-8	7.0 +/- 2.1	-11.9 +/- 1.6	0.1 +/- 1.2
9-10	9.0 +/-1.9	-11.5 +/- 1.3	7.2 +/- 2.7
11-12	10 +/- 3.8	-9 +/-1.4	10.9 +/- 2.9
13-14	12.5 +/- 4.4	-9.5 +/- 1.6	12.3 +/- 6.4
15-16	11.1 +/- 1.0	-3.0_+/- 1.5	10.3 +/- 3.3
17-18	17.2 +/- 3.6	8.0 +/- 1.3	13.3 +/- 3.4

As can be seen, at the end of a day 17 after a 7 day subcutaneous regime at 10 mg/kg/day, animals receiving the FC-OB protein recovered to 8% of their body weight. Animals receiving dosages of 1 mg/kg/day after a 7 day subcutaneous regime returned to 6.4% of body weight after 12 days.

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These studies also show that during recovery periods from day 7 to day 22, after the last injection at day 7, body weight recovery is slower in the Fc-OB treated C57 mice that with the OB treated mice. This suggests that the Fc-OB protein is not cleared as quickly as OB protein thereby causing the extended weight loss effect.

EXAMPLE 3: Dose Response of CF7 Mice Treated with Fc-OB Fusion Protein

An additional study demonstrated that there was a dose response to continuous administration of Fc-OB protein. In this study, obese CF7 mice, weighing

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35-40 g were administered recombinant human Fc-OB protein using methods similar to the above example. The results are set forth in Table 3, below, (with % body weight lost as compared to baseline, measured as above):

Table 3

Dose Response With Continuous Administration

Dose	Time	% Reduction in Body Weight
0.25 mg/kg/day	Day 5	4
0.5 mg/kg/day	Day 5	12
1 mg/kg/day	Day 5	16

10 As can be seen, increasing the dose from 0.25 mg/kg/day to 1 mg/kg/day increased the weight lost from 4% to 16%. It is also noteworthy that at day 5, the 1 mg/kg/day dosage resulted in a 16% reduction in body weight. These studies also showed slow weight recovery 15 rates to 0% suggesting that the Fc-OB protein is not quickly cleared thereby causing the extended weight loss effect.

EXAMPLE 4: Pharmacokinetics of recombinant human Fc-OB in CD-1 Mice and Dogs

This study demonstrated the pharmacokinetic properties of recombinant human met Fc-OB protein in CD-1 mice and dogs. Following intravenous or subcutaneous dosing at 1 mg/kg/day, serum concentrations of recombinant human met Fc-OB protein and human met OB protein were determined by an enzyme-linked immunosorbent assav (ELISA).

In both species, an increase in exposure, as quantified by higher peak serum concentrations and larger ar as under-the-serum-concentration-curve (AUC), was observed when compared to recombinant met-human OB

PCT/US97/23183

-36-

protein. Fc-OB has lower systemic clearance than recombinant met-human OB protein. This is seen in the lower clearance and longer half-life of Fc-OB over OB protein. The increase in size causes not only an increase in protein stability, but also a decrease in the efficiency of renal clearance. As a result, Fc-OB is cleared slower from the systemic circulation. The increases in peak time, peak serum concentrations and AUC for Fc-OB protein are consistent with lower clearance. Fc-OB protein will yield substantially higher systemic exposure when compared to OB protein.

Table 4

Pharmacokinetic Properties

Species	CD-1	Mice	CD-1	Mice	Beagle Dogs		
Route of		venous	Subcut	aneous	Subcutaneous		
	ОВ	Fc-OB protein	OB protein	Fc-OB protein	OB protein	Fc-OB protein	
Dose Level (mg/kg)	1	1	1	1	0.5	0.5	
Peak Time (h)			0.14	6	2.8	8	
Peak Serum Concentration (ng/mL)			1520	7550	300	1120	
AUC (ngeh/mL)	1470	366000	1230	132000	2200-	52500	
Half-life (h)	0.491	21.4	0.388		2.13	22.9	
Clearance (mL/h/kg)	681	2.73) (a) 	19-19-1		

EXAMPLE 5:

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This example demonstrates that in normal mice

which are not obese and do not have elevated blood lipid
levels, administration of human recombinant Fc-OB

protein results in a lowering of cholesterol, glucose and triglyceride levels. In addition, this example demonstrates that these levels remain low over a three day recovery period.

Normal CD1 mice were administered recombinant human Fc-OB protein via subcutaneous injections. Blood samples were taken 24 hours after day 23, the last day of injection. As discussed above, the animals lost weight at the dosages administered. As shown in Table 5, the mice had substantial reduction of serum cholesterol, glucose and triglycerides in a dosedependent fashion when compared to controls:

Table 5

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Dose	Glucose	Cholesterol	Triglycerides
PBS	232.6 +/- 15.1	67.8 +/- 3.6	52.6 +/- 3.7
1 mg/kg/day	225.8 +/- 29.1	54 +/- 5.6	43 +/- 8.7
10 mg/kg/day	193.2 +/- 21.4	53.4 +/- 5.7	38 +/- 11
1 mg/kg every 2 days	242.0 +/- 9.3	52.6 +/- 4.4	40.8 +/- 7.2
10 mg/kg every 2 days	197.4 +/- 27.9	51.4 +/- 5.9	29.8 +/- 6.3
1 mg/kg every 3 days	244.8 +/- 19.5	60.8 +/- 7.3	54 +/- 7.1
10 mg/kg every 3 days	188 +/- 31.2	52.2 +/- 6.9	26.2 +/- 10.7

These data demonstrate that the Fc-OB protein, or analogs or derivatives thereof, are effective blood lipid lowering agents.

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EXAMPLE 6:

A obese human patient is administered human Fc-OB protein, or analog or derivative for the purpose of weight reduction. The obese patient also has elevated

levels of blood lipids, including elevated levels of cholesterol, above 200 mg/100 ml. The patient attains a satisfactory weight reduction over the course of FC-OB therapy. A maintenance dose of FC-OB protein or analog or derivative is administered to the non-obese patient to maintain lowered blood lipid levels, including lowered cholesterol levels, below 200 mg/100 ml. The dose administered is insufficient to result in further weight loss. Administration is chronic. Levels of circulating FC-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

15 EXAMPLE 7:

A non-obese human patient undergoes coronary bypass surgery or other invasive treatment to alleviate advanced stages arterial plaque formation. After the surgery, the patient is administered a maintenance dose of Fc-OB protein or analog or derivative in order to prevent the re-formation of arterial plaque. The dose administered is insufficient to result in weight loss. Administration is chronic. Levels of circulating Fc-OB protein or analog or derivative may be monitored using 25 a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

EXAMPLE 8:

A non-obese human patient experiences

hypertension due to restricted blood flow from clogged arteries. The patient is administered a dose of Fc-OB protein, or analog or derivative thereof sufficient to reduce arterial plaque resulting in clogged arteries. Thereafter, the patient is monitored for further arterial plaque formation, and hypertension. If the condition re-appears, the patient is re-administered an

PCT/US97/23183

effective amount of Fc-OB protein, analog or derivative sufficient to restore blood flow, yet insufficient to result in weight loss. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the Fc-OB protein (or other antigenic source if applicable).

EXAMPLE 9:

A human patient experiences gall stones. 10 Either the gall stones are not removed and the formation of additional gall stones is sought to be avoided, or the gall stones are removed but the gall bladder remains (as, for example, using laser or ultrasonic surgery) and the formation of additional gall stones is sought to be avoided. The patient is administered an effective amount of Fc-OB protein, analog or derivative thereof to result in prevention of accumulation of additional gall stones or re-accumulation of gall stones. Levels of circulating Fc-OB protein or analog or derivative may be 20 monitored using a diagnostic kit, such as an antibody assay against the Fc-OB protein (or other antigenic source if applicable).

EXAMPLE 10:

25 A diabetic human patient desires to use decreased dosages of insulin for treatment of diabetes. The patient is administered an effective amount of Fc-OB protein, analog or derivative thereof to result in an increase in lean tissue mass. The patient's sensitivity 30 to insulin increases, and the dosage of insulin necessary to alleviate symptoms of diabetes is decreased, either in terms of a decrease in the units of insulin n eded, or in terms of a decrease in the number of injections of insulin needed per day. Levels of 35 circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

5 EXAMPLE 11:

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A non-obese human patient desires an increase in lean tissue mass for therapeutic purposes, such as recovery from illness which depleted lean tissue mass. The patient is administered an effective amount of Fc-OB protein, analog or derivative thereof to result in the desired increase in lean tissue mass. Increase in lean tissue mass is monitored using DEXA scanning. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

MATERIALS AND METHODS

- 20 <u>Animals</u>. Wild type CD1 mice and (+/+) C57B16 mice were used for the above examples. The age of the mice at the initial time point was 8 weeks, and the animals were weight stabilized.
- 25 <u>Feeding and Weight Measurement.</u> Mice were given ground rodent chow (PMI Feeds, Inc.) in powdered food feeders (Allentown Caging and Equipment) which allowed a more accurate and sensitive measurement than use of regular block chow. Weight was measured at the 30 same time each day (2:00 p.m.), for the desired period. Body weight on the day prior to the injection was defined as baseline weight. The mice used weighed 18-22 grams.
- 35 <u>Housing</u>. Mice were single-housed, and maintained under humane conditions.

PCT/US97/23183 W-O 98/28427

-41-

Administration of Protein or Vehicle. Protein (as described below) or vehicle (phosphate buffered saline, pH 7.4) were administered by subcutaneous injections or intravenously.

Controls. Control animals were those who were injected with the vehicle alone without either Fc-OR fusion protein or OB protein added to the vehicle.

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Protein. Sequence ID. Nos. 1, 2 and 3 set forth murine recombinant OB DNA and protein (Figure 1). and Sequence ID. Nos. 4, 5 and 6 set forth an analog recombinant human OB DNA and protein (Figure 2). As noted 15 above recombinant human OB protein as in SEO. ID., NO. 6 has a lysine residue at position 35 and an isoleucine residue at position 74. Furthermore, the recombinant human protein set forth in Zhang et al., Nature, supra, and PCT publication WO 96/05309 (12/22/96) (both

20 incorporated by reference including figures), and the murine and human analog recombinant proteins of Figures 1 and 2 are illustrative of the OB protein which may be used in forming the Fc-OB fusion protein of the present methods of treatment and manufacture of a medicament. Other OB or 25 Fc proteins or analogs or derivatives thereof may also be used to form the Fc-OB fusion protein.

Herein, the first amino acid of the amino acid sequence for recombinant OB protein is referred to as +1, and is valine, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 146 (cysteine) (see Figures 1 and 2). The first amino acid sequence for recombinant human Fc-OB protein of Figure 3 is referred to as +1, and is glutamate, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 378 (cysteine). The first amino acid sequence for the recombinant human Fc-OB protein variant

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of Figure 4 is referred to as +1, and is glutamate, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 378 (cysteine). The first amino acid sequence for the recombinant human 5 Fc-OB protein variant of Figure 5 is referred to as +1, and is aspartic acid, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 373 (cysteine). The first amino acid sequence for the recombinant human Fc-OB protein variant of Figure 6 is 10 referred to as +1, and is aspartic acid, and the amino acid at position -1 is methionine. The C-terminal amino acid is number is 373 (cysteine).

Expression Vector and Host Strain

The plasmid expression vector used is pAMG21

(ATCC accession number 98113), which is a derivative of pCFM1656 (ATCC accession number 69576) and contains appropriate restriction sites for insertion of genes downstream from the lux PR promoter (see US Patent No. 5,169,318 for a description of the lux expression system). The Fc-OB DNA, described below and shown in Figures 3-6, was created and ligated into the expression vector pAMG21 linearized with restriction endonucleases NdeI and BamHI and transformed into the E. coli host strain, FM5. E. coli FM5 cells were derived at Amgen

- Inc., Thousand Oaks, CA from E. <u>coli</u> K-12 strain (Bachmann, et al., Bacterial. Rev. <u>40</u>: 116-167 (1976)) and contain the integrated lambda phage repressor gene, cI857 (Sussman et al., C. R. Acad. Sci. <u>254</u>: 1517-1579
- 30 (1962)). Vector production, cell transformation, and colony selection were performed by standard methods, (e.g., Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2d Edition, Cold Spring Harbor Laboratory Pr ss, Cold Spring Harbor, NY.) Host cells were grown in LB media.

-43-

Fc-OB DNA Construction

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The plasmid pFc-A3 (described below) served as the source of sequence for human immunoglobulin IgG-1 heavy chain from amino acid number 99 (Glu) to the natural carboxyl terminus. The human IgG-1 sequence can be obtained from Genebank (P01857).

The human OB sequence is disclosed above as well as Zhang et al., Nature, <u>supra</u>, and PCT publication WO 96/05309 both incorporated by reference including drawings. The OB DNA was ligated into the expression vector pCFM1656 linearized with restriction

endonucleases XbaI and BamHI using standard cloning procedures, e.g., Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2d Edition, Cold Spring Harbor Laboratory Press, Cold Spting Harbor, N.Y.. The plasmid

pCFM1656 carrying the OB DNA sequence served as the source of sequence for the recombinant human OB gene.

The genetic fusing of these two sequences was carried out by the method of PCR overlap extension (Ho.

- 20 S.N., et al., Site Directed Mutagenesis By Overlap
 Extension Using The Polymerase Chain Reaction, Gene
 77:51-59(1989)). The product of the PCR was cleaved
 with restriction endonuclease NdeI to create a
 5'-cohesive end and with restriction endonuclease BamHI
- 25 to create a 3'-cohesive terminus. The vector, pAMG21, was similarly cleaved. A ligation was performed with the fusion fragment and the linearized vector. Ligated DNA was transformed by electroporation into the E. coli host strain. Clones surviving on kanamycin (50µg/ml)
- 30 selection agar plates were checked for expression of Fc-OB-sized protein. Plasmid from individual clones was isolated and the sequence of the gene coding region verified.

When additional modifications of the Fc-OB 5 gene were desired, the PCR technique was used again to engineer the changes. Two sets of changes were

performed at the N-terminus of the Fc portion of the fusion protein (SEO, ID, No. 9) to create the variants SEO. ID. NOS. 12 and 15. Another variant was constructed to introduce four amino acid substitutions to ablate the Fc-receptor binding site (leucine at position 15 substituted with glutamate), and the complement (Clg) binding site (glutamate at position 98 substituted with alanine, lysine at position 100 substituted with alanine, and lysine at position 102 10 substituted with alanine (See, Xin Xiao Zheng et. al, J. Immunol. 154:5590-5600 (1995)). The template for this construct was Seq. ID. No. 15 and the resulting variant was SEO. ID. Nos. 18.

15 pFC-A3 Vector Construction

A plasmid, pFc-A3, containing the region encoding the Fc portion of human immunoglobulin IgG-1 heavy chain (See Ellison, J. W. et. al, Nucleic Acids Res. 10:4071-4079 (1982)), from the first amino acid 20 Glu-99 of the hinge domain to the carboxyl terminus plus a 5'-NotI fusion site and 3'-SalI and XbaI sites, was made by PCR amplification of the human spleen cDNA library. PCR reactions were in a final volume of 100 ml and employed 2 units of Vent DNA polymerase in 20 mM Tris-HCl (pH 8.8), 10 mM KCl, 10 mM (NH4)2SO4, 2 mM MgSO4, 0.1% Triton X-100 with 400 mM each dNTP and 1 ng of the cDNA library to be amplified together with 1 uM of each primer. Reactions were initiated by denaturation at 95 °C for 2 min, followed by 30 cycles of 95 °C for 30 30 s, 55 °C for 30 s, and 73 °C for 2 min. The 5'primer incorporated a NotI site immediately 5' to the first residue (Glu-99) of the hinge domain of IgG-1. The 3'-primer incorporated SalI and XbaI sites. The 717 base pair PCR product was digested with NotI and SalI, 35 the resulting DNA fragment was isolated by

electrophoresis through 1% agarose and purified and

PCT/IIS97/23183 WO 98/28427

cloned into NotI, SalI-digested pBluescript II KS vector (Stratagene). The insert in the resulting plasmid, pFc-A3, was sequenced to confirm the fidelity of the PCR

-45-

reaction.

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Methods for Production

The methods below for production have been used to produce biologically active recombinant methionyl murine or human analog OB protein and Fc-OB fusion proteins. Similar methods may be used to prepare biologically active methionyl human OB protein.

Fermentation Process

A batch fermentation process was used. Media compositions are set forth below. 15

A portion of the media consisting of primarily nitrogen sources was sterilized (by raising temperature to 120~123°C for 25-35 minutes) in the fermentation vessel. Upon cooling, carbon, magnesium, phosphate, and trace metal sources were added aseptically. An overnight culture of the above recombinant murine protein-producing bacteria of 500 mL (grown in LB broth) was added to the fermentor. When the culture optical density (measured at 600 nm as an indicator for cell

- density) reached 15~25 absorption units, an autoinducer 25 solution (0.5 mg/mL homoserine lactone) was added (1 mL/L) to the culture to induce the recombinant gene expression. The fermentation process was allowed to continue for additional 10 to 16 hours, followed by 30 harvesting the broth by centrifugation.

PCT/IIS97/23183 WiD 98/28427

-46-

Media Composition:

Batch: 34 g/L Yeast extract 78 g/L Sov peptone 0.9 g/L Potassium chloride 5.0 q/L Hexaphos 5 1.7 g/L Citric acid 120 g/L Glycerol 0.5 g/L MgSO4 · 7H2O 0.2 mL/L Trace Metal Solution 0.5 mL/L P2000 Antifoam 10

Trace Metal Solution:

incorporated by reference.

25

Ferric Chloride (FeCl3 · 6H2O): 27 g/L Zinc Chloride (ZnCl2·4H2O): 2 g/L 15 Cobalt Chloride (CoCl2.6H2O): 2 g/L Sodium Molybdate (NaMoO4 · 2H2O): 2 g/L Calcium Chloride (CaCl2 · 2H2O): 1 g/L Cupric Sulfate (CuSO4.5H2O): 1.9 g/L Boric Acid (H3BO3): 0.5 g/L 20 Manganese Chloride (MnCl2 · 4H2O): 1.6 g/L Sodium Citrate dihydrate: 73.5 g/L

Purification Process for Human Fc-OB Fusion Protein

Purification for human Fc-OB fusion protein was accomplished by the steps below (unless otherwise noted, the following steps were performed at 4'C). Purification for murine and human OB protein is 30 disclosed in PCT publication WO 96/05309, supra, herein

1. Cell paste. E. coli cell paste was suspended in 5 times volumes of distilled water. The cells in the water were further broken by two passes 10

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through a microfluidizer. The broken cells were centrifuged at 4.2k rpm for 1 hour in a Beckman JB-6 centrifuge with a J5-4.2 rotor.

- 2. Inclusion body wash. The supernatant from above was removed and the pellet was resuspended with five volumes of distilled water. The mixture was centrifued as in step 1.
- 3. Solubilization. The pellet was solubilized with 10 volumes of 50 mM tris, pH 8.5, 8 M guanidine hydrochloride, 10 mM dithiothreitol and stirred for one hour at room temperature. The solution is made 40 mM cystamine dihydrochloride and stirred for one hour.
- 4. The solution from step 3 is added to 20 to 30 volumes of the following refold solution: 50 mM tris, pH 8.5, 0.8 M arginine, 2 M urea, and 4 mM cysteine. The refold is stirred for 16 hours at 8°C.
- 5. Buffer exchange. The solution from step 4 is concentrated and diafiltered into 10 mM tris, pH 8.5.
- 6. Acid precipitation. The solution from step 20 5 is adjusted to pH 4.75 with 50% glacial acid and incubated for 30 minutes at room temperature. The solution is filtered.
- Cation exchange chromatography. The solution from step 6 is adjusted to pH 7.0 and loaded onto a CM Sepharose Fast Flow column at 10°C. A twenty column volume gradient is done at 10 mM phosphate, pH 7.0. 0 to 0.1 M NaCl.
- Anion exchange chromatography. The CM elution pool from step 7 is diluted 5 fold with 5 mM
 tris, pH 7.5 and loaded onto a Q Sepharose Fast Flow at 10°C. A 20 column volume gradient is done at 10 mM tris, pH 7.5, 0 to 0.2M Nacl.
- Hydrophobic interaction chromatography. The Q sepharose pool is made 0.75M ammonium sulfate and
 loaded on a m thyl Macroprep hydrophobic interaction

-48-

column at room temperature. A 20 column volume gradient is done at 10 mM phosphate, pH 7.0, 0.75M to 0M ammonium sulfate.

10. Buffer exchange. The pool from step 9 is 5 concentrated as necessary and dialyzed against PBS buffer.

While the present invention has been described in terms of preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of the invention as claimed.

- 49 -

SECUENCE LISTING

(1) GENERAL INFORMATION:

- (1) APPLICANT: Mann, Michael B. Hecht. Randy I.
- (ii) TITLE OF INVENTION: OB FUSION PROTEIN COMPOSITIONS AND
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Amgen Inc.
 - (B) STREET: 1840 DeHavilland Drive
 - (C) CITY: Thousand Oaks
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 91320-1789
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/770,973
 - (B) FILING DATE: 20-DEC-1996
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Knight, Matthew W.
 - (B) REGISTRATION NUMBER: 36,846
 - (C) REFERENCE/DOCKET NUMBER: A-416

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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 491 base pairs (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- .

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
 - (B) LOCATION: 41
 - (D) OTHER INFORMATION: /note: "Met = ATG"

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60

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- 50 -

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- 51 -

Pro Glu Cys 145

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    Ile Leu Ser Leu Ser Lys Met Asp Gln Thr Leu Ala Val Tyr Gln Gln
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- 52 -

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(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: CDNA	
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WO 98/28427	PCT/US97/23183

- 53 -

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(xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q II	NO:	6:							
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Cys	His	Leu	Pro	Trp	Ala	Ser	Gly	Leu 105	Glu	Thr	Leu	Asp	Ser 110		Glγ	

- 54 -

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	Arg	Leu 130	Gln	Gly	Ser	Leu	Gln 135	Asp	Met	Leu	Trp	Gln 140	Leu	Asp	Leu	Ser
	Pro 145	Gly	Cys													
2)	2) INFORMATION FOR SEQ ID NO:7:															
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1150 base pairs (B) TYPE: nucleic acid (C) STRANDENDESS: double (D) TOPOLOGY: linear															
	(ii)	MOLI	ECUL	E TY	PE: «	DNA										
	<pre>(ix) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 4 (D) OTHER INFORMATION: /note= 'Met = ATG'</pre>															
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:																
27	ATATIGNAC CCALATOTTIC TOLCALAROT CACACITACCO CACCOTTCCCC ACCACITACA															

CATATGGAAC CCAAATCTTG TGACAAAACT CACACATGCC CACCGTGCCC AGCACCTGAA 60 CTCCTGGGGG GACCGTCAGT CTTCCTCTTC CCCCCAAAAC CCAAGGACAC CCTCATGATC 120 TCCCGGACCC CTGAGGTCAC ATGCGTGGTG GTGGACGTGA GCCACGAAGA CCCTGAGGTC 180 AAGTTCAACT GGTACGTGGA CGGCGTGGAG GTGCATAATG CCAAGACAAA GCCGCGGGAG 240 GAGCAGTACA ACAGCACGTA CCGTGTGGTC AGCGTCCTCA CCGTCCTGCA CCAGGACTGG 300 CTGAATGGCA AGGAGTACAA GTGCAAGGTC TCCAACAAAG CCCTCCCAGC CCCCATCGAG 360 AAAACCATCT CCAAAGCCAA AGGGCAGCCC CGAGAACCAC AGGTGTACAC CCTGCCCCCA 420 TCCCGGGATG AGCTGACCAA GAACCAGGTC AGCCTGACCT GCCTGGTCAA AGGCTTCTAT 480 CCCAGCGACA TCGCCGTGGA GTGGGAGAGC AATGGGCAGC CGGAGAACAA CTACAAGACC 540 ACCCCTCCCG TGCTGGACTC CGACGGCTCC TTCTTCCTCT ACAGCAAGCT CACCGTGGAC 600 AAGAGCAGGT GGCAGCAGGG GAACGTCTTC TCATGCTCCG TGATGCATGA GGCTCTGCAC 660 AACCACTACA CGCAGAAGAG CCTCTCCCTG TCTCCGGGTA AAGTACCGAT CCAGAAAGTT 720

- 55 -

CAGGACGACA	CCAAAACCTT	AATTAAAACG	ATCGTTACGC	GTATCAACGA	CATCAGTCAC	780
ACCCAGTCGG	TGAGCTCTAA	ACAGAAAGTT	ACAGGCCTGG	ACTTCATCC	GGGTCTGCAC	840
CCGATCCTGA	CCTTGTCCAA	AATGGACCAG	ACCCTGGCTG	TATACCAGCA	GATCTTAACC	900
TCCATGCCGT	CCCGTAACGT	TATCCAGATO	TCTAACGACC	TOGAGAACOT	TCGCGACCTG	960
CTGCACGTGC	TGGCATTCTC	CAAATCCTGC	CACCTGCCAT	GGGCTTCAGG	TCTTGAGACT	1020
CTGGACTCTC	TGGGCGGGGT	CCTGGAAGCA	TCCGGTTACA	GCACCGAAGT	TGTTGCTCTG	1080
TCCCGTCTGC	AGGGTTCCCT	TCAGGACATG	CTTTGGCAGC	TGGACCTGTC	TCCGGGTTGT	1140
TAATGGATCC						1150

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1150 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GTATACCTTG	GGTTTAGAAC	ACTGTTTTGA	GTGTGTACGG	GTGGCACGGG	TCGTGGACTT	60
GAGGACCCCC	CTGGCAGTCA	GAAGGAGAAG	GGGGGTTTTG	GCTTCCTGTG	GGAGTACTAG	120
AGGCCTGGG	GACTCCAGTG	TACGCACCAC	CACCTGCACT	CGCTGCTTCT	GGGACTCCAG	180
TTCAAGTTGA	CCATGCACCT	GCCGCACCTC	CACGTATTAC	GGTTCTGTTT	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	240
CTCGTCATGT	TGTCGTGCAT	GGCACACCAG	TCGCAGGAGT	GGCAGGACGT	GGTCCTGACC	300
GACTTACCGT	TCCTCATGTT	CACGTTCCAG	AGGTTGTTTC	GGGAGGGTCG	GGGGTAGCTC	360
TTTTGGTAGA	COTTTCCCTT	TCCCGTCGGG	CCTCTTCCTC	TCCACATGTG	GGACGGGGT	420
AGGGCCCTAC	TCGACTGGTT	CTTGGTCCAG	TCGGACTGGA	CGGACCAGTT	TCCGAAGATA	480
GGGTCGCTGT	AGCGGCACCT	CACCCTCTCG	TTACCCGTCG	GCCTCTTGTT	GATGTTCTGG	540
TGCGGAGGGC	ACGACCTGAG	GCTGCCGAGG	AAGAAGGAGA	TGTCGTTCGA	GTGGCACCTG	600
TTCTCGTCCA	CCGTCGTCCC	CTTGCAGAAG	AGTACGAGGC	ACTACGTACT	CCGAGACGTG	660
TTGGTGATGT	GCGTCTTCTC	GGAGAGGGAC	AGAGGCCCAT	TTCATGGCTA	GGTCTTTCAA	720

WO 98/28427		PCT/US97/23183
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78C

- 56 -

GTCCTGCTGT GGTTTTGGAA TTAATTTTGC TAGCAATGCG CATAGTTGCT GTAGTCAGTG

TGGGTCAGCC ACTCGAGATT TGTCTTTCAA TGTCCGGACC TGAAGTAGGG CCCAGACGTG	840
GGCTAGGACT GGAACAGGTT TTACCTGGTC TGGGACCGAC ATATGGTCGT CTAGAATTGG	900
AGGTACGGCA GGGCATTGCA ATAGGTCTAG AGATTGCTGG AGCTCTTGGA AGCGCTGGAC	960
GACGTGCACG ACCGTAAGAG GTTTAGGACG GTGGACGGTA CCCGAAGTCC AGAACTCTGA	1020
GACCTGAGAG ACCCGCCCCA GGACCTTCGT AGGCCAATGT CGTGGCTTCA ACAACGAGAC	1080
AGGGCAGACG TCCCAAGGGA AGTCCTGTAC GAAACCGTCG ACCTGGACAG AGGCCCAACA	1140
ATTACCTAGG	1150
(2) INFORMATION FOR SEQ ID NO:9: (i) SEQUENCE CHARACTERISTICS: (A) LENTH: 379 amino acids	
(B) TYPE: amino acid (C) STRANDEDNESS: unknown (D) TOPOLOGY: unknown	
(ii) MOLECULE TYPE: protein	
(ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1 (D) OTHER INFORMATION: /note= 'Met (ATG) starts at -1'	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
Met Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys F .1 5 10	TO
Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro I 20 25 30	ys
Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys V 35 40	7 a l
Val Val Asp Val Ser His Glu Asp Pr Glu Val Lys Phe Asn Trp 5 50 60	lyr
Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu	Glu

- 57 -

Gln	Tyr	Asn		Thr 85	Tyr	Arg	Val	Val	Ser 90	Val	Leu	Thr		Leu 95	Hıs
Gln	Asp	Trp	Leu 100	Asn	Gly	Lys	Glu	Tyr 105	Lys	Cys	Lys	Val	Ser 110	Asn	Lys
λlα	Leu	Pro 115	Ala	Pro	Ile	Glu	Lys 120	Thr	Ile	Ser	Lys	Ala 125	Lys	Gly	Gln
Pro	Arg 130	Glu	Pro	Gln	Val	Tyr 135	Thr	Leu	Pro	Pro	Ser 140	Arg	As p	Glu	Leu
Thr 145	Lys	As n	Gln	Val	Ser 150	Leu	Thr	Cys	Leu	Val 155	Lys	Gly	Phe	Tyr	Pro 160
Ser	Asp	Ile	Ala	Val 165	Glu	Trp	Glu	Ser	Asn 170	Gly	Gln	Pro	Glu	Asn 175	As n
Tyr	Lys	Thr	Thr 180	Pro	Pro	Val	Leu	Asp 185	Ser	Asp	Gly	Ser	Phe 190	Phe	Leu
Tyr	Ser	Lys 195	Leu	Thr	Val	Asp	Lys 200	Ser	Arg	Trp	Gln	Gln 205	Gly	As n	Val
Phe	Ser 210	Cys	Ser	Val	Met	His 215	Glu	Ala	Leu	His	Asn 220	His	Tyr	Thr	Gln
Lys 225		Leu	Ser	Leu	Ser 230	Pro	Gly	Lys	Val	Pro 235	Ile	Gln	Lys	Val	Gln 240
225					230		Gly Lys	-		235			•		240
Asp	Asp	Thr	Lys	Thr 245	230 Leu	Ile	_	Thr	Ile 250	235 Val	Thr	Arg	Ile	Asn 255 Gly	240 Asp
Asp	Asp Ser	Thr	Thr 260	Thr 245 Gln	230 Leu Ser	Ile Val	Lys	Thr Ser 265	Ile 250 Lys	Val Gln	Thr Lys	Arg Val	Ile Thr 270	Asn 255 Gly	240 Asp Leu
225 Asp Ile	Asp Ser	Thr His 275	Thr 260	Thr 245 Gln Gly	230 Leu Ser	Ile Val	Lys Ser Pro 280	Thr Ser 265	Ile 250 Lys	Val Gln Thr	Thr Lys	Arg Val Ser 285	Thr 270 Lys	Asn 255 Gly Met	240 Asp Leu Asp
Asp Ile	Asp Ser Phe Thr 290	Thr His 275	Thr 260 Pro	Thr 245 Gln Gly Val	230 Leu Ser Leu	Val His Gln 295	Lys Ser Pro 280	Thr Ser 265 Ile	Ile 250 Lys Leu	Val Gln Thr	Thr Lys Leu Ser 300	Arg Val Ser 285	Thr 270 Lys	Asn 255 Gly Met	240 Asp Leu Asp
225 Asp Ile Asp Glr Ass	Asp Ser Phe Thr 290	Thr His 275 Let	Thr 260 Pro	Thr 245 Gln Gly Val	230 Leu Ser Leu Tyr Ser 310	Val His Glm 295	Lys Ser Pro 280	Thr Ser 265 Ile	Ile 250 Lys Leu Leu	Val Gln Thr Thr Asn 315	Lys Leu Ser 300	Arg Val Ser 285 Met	Thr 270 Lys Pro	Asn 255 Gly Met Ser	240 Amp Leu Amp Arg Leu 320 Gly
2255 Asp Ile Asp Glr Ass Strict A	Asp Ser Pho Thr 290 1 Val	Thr His : Ile 275 : Len	Lys Thr 260 Pro	Thr 245 Gln Gly Val Val 329	230 Leu Ser Leu Tyr	Ile Val His Glm 295 Asr	Lys Ser Pro 280	Thr Ser 265 Ile Ile Leu Cyr	Ile 250 Lys Leu Leu Glu His His 330	Val Gln Thr Thr Asn 315	Leu Ser 300	Arg Val Ser 285 Met	The Thr 270 Lys Pro	Asn 255 Gly Met Ser Leu Ser 335	Amp Leu Amp Arg Leu 320

- 58 -

Met Leu Trp Gln Leu Asp Leu Ser Pro Gly Cys 370 375

- (2) INFORMATION FOR SEQ ID NO:10:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1150 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 - . . .
 - (ii) MOLECULE TYPE: cDNA
 - (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION: 4
 - (D) OTHER INFORMATION: /note= "Met = ATG"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CATATGGAAC CAAAATCTGC TGACAAAACT CACACATGTC CACCTTGTCC AGCTCCGGAA 60 CTCCTGGGGG GTCCTTCAGT CTTCCTCTTC CCCCCAAAAC CCAAGGACAC CCTCATGATC 120 TOCCGGACCO CTGAGGTCAC ATGCGTGGTG GTGGACGTGA GCCACGAAGA CCCTGAGGTC 180 AAGTTCAACT GGTACGTGGA CGGCGTGGAG GTGCATAATG CCAAGACAAA GCCGCGGGAG 240 GAGCAGTACA ACAGCACGTA CCGTGTGGTC AGCGTCCTCA CCGTCCTGCA CCAGGACTGG 300 CTGAATGGCA AGGAGTACAA GTGCAAGGTC TCCAACAAAG CCCTCCCAGC CCCCATCGAG 360 ANARCRATCT CCARAGCCAR AGGGCAGCCC CGAGAACCAC AGGTGTACAC CCTGCCCCCA 420 TCCCGGGATG AGCTGACCAA GAACCAGGTC AGCCTGACCT GCCTGGTCAA AGGCTTCTAT 480 CCCAGCGACA TCGCCGTGGA GTGGGAGAGC AATGGGCAGC CGGAGAACAA CTACAAGACC 540 ACCCCTCCCG TGCTGGACTC CGACGGCTCC TTCTTCCTCT ACAGCAAGCT CACCGTGGAC 600 ANGAGEAGGT GGEAGEAGGG GAACGTETTE TEATGETEEG TGATGEATGA GGETETGEAE 660 AACCACTACA CGCAGAAGAG CCTCTCCCTG TCTCCGGGTA AAGTACCGAT CCAGAAAGTT 720 CAGGACGACA CCAAAACCTT AATTAAAACG ATCGTTACGC GTATCAACGA CATCAGTCAC 780 ACCCAGTOGG TGAGCTCTAA ACAGAAAGTT ACAGGCCTGG ACTTCATCCC GGGTCTGCAC 840 CCGATCCTGA CCTTGTCCAA AATGGACCAG ACCCTGGCTG TATACCAGCA GATCTTAACC 900

- 59 -

TCCATGCCGT	CCCGTAACGT	TATCCAGATO	TCTAACGACC	TCGAGAACCT	TCGCGACCTG	960
CTGCACGTGC	TGGCATTCTC	CAAATCCTGC	CACCTGCCAT	GGGCTTCAGG	TCTTGAGACT	1020
CTGGACTCTC	TGGGCGGGGT	CCTGGAAGCA	TCCGGTTACA	GCACCGAAGT	TGTTGCTCTG	1083
TCCCGTCTGC	AGGGTTCCCT	TCAGGACATG	CTTTGGCAGC	TGGACCTGTC	TCCGGGTTGT	1140
TAATGGATCC						1150

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1150 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GTATACCTTG GTTTTAGACG ACTGTTTTGA GTGTGTACAG GTGGAACAGG TCGAGGCCTT 60 GAGGACCCCC CAGGAAGTCA GAAGGAGAAG GGGGGTTTTG GGTTCCTGTG GGAGTACTAG 120 AGGGCCTGGG GACTCCAGTG TACGCACCAC CACCTGCACT CGGTGCTTCT GGGACTCCAG 180 TTCAAGTTGA CCATGCACCT GCCGCACCTC CACGTATTAC GGTTCTGTTT CGGCGCCCTC 240 CTCGTCATGT TGTCGTGCAT GGCACACCAG TCGCAGGAGT GGCAGGACGT GGTCCTGACC 300 GACTTACCGT TCCTCATGTT CACGTTCCAG AGGTTGTTTC GGGAGGGTCG GGGGTAGCTC 360 TITTGGTAGA GGTTTCGGTT TCCCGTCGGG GCTCTTGGTG TCCACATGTG GGACGGGGGT 420 AGGGCCCTAC TEGACTGGTT CTTGGTCCAG TEGGACTGGA CGGACCAGTT TEEGAAGATA 480 GGGTCGCTGT AGCGGCACCT CACCCTCTCG TTACCCGTCG GCCTCTTGTT GATGTTCTGG 540 TGCGGAGGGC ACGACCTGAG GCTGCCGAGG AAGAAGGAGA TGTCGTTCGA GTGGCACCTG 600 TTCTCGTCCA CCGTCGTCCC CTTGCAGAAG AGTACGAGGC ACTACGTACT CCGAGACGTG 660 TTGGTGATGT GCGTCTTCTC GGAGAGGGAC AGAGGCCCAT TTCATGGCTA GGTCTTTCAA 720 GTCCTGCTGT GGTTTTGGAA TTAATTTTGC TAGCAATGCG CATAGTTGCT GTAGTCAGTG 780 TGGGTCAGCC ACTCGAGATT TGTCTTTCAA TGTCCGGACC TGAAGTAGGG CCCAGACGTG 840

WO 98/28427	PCT/US97/23183

	60	-
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GGCTAGGAC	T GG.	AACA	GTT	TTAC	CTGC	TC T	rggg	ACCG/	AC A	TATG	GTCG	CT	AGAA	TTGG		900
AGGTACGGC.	λ GG	GCAT	rgca	ATA	GTCI	rag i	AGAT	IGCT	GG A	CTC	TTGG	A AG	CGCT	GGAC		960
GACGTGCAC	G AC	CGTA	AGAG	GTT:	PAGGA	LCG (STGG	ACGG	ra c	CCGA	AGTC	C AG	AACT	CTGA		1020
GACCTGAGA	G AC	cccc	CCCA	GGA	CTTC	GT .	AGGC	CAAT	ST C	STCC	CTTC	A AC	AACG	AGAC		1080
AGGGCAGAC	G TC	CCAA	GGGA	AGT	CTG	rac (GAAA	CCGT	CG A	crc	GACA	G AG	GCCC	AACA		1140
ATTACCTAG	G															1150
(2) INFOR	SEQU (A) (B) (C)	ENCE LEN TYP STR	CHAIGTH: E: as ANDES	379 sino	ERIST amin acid	rics no a 1 nkno	cids									
(ii)	MOLE	CULE	TYP	E: p	rote	in										
(ix)	(A) (B) (D)	LOC	E/KE ATIO ER I	N: 1 NFOR	MATI	OIN :				(ATG) st	arte	at	-1•		
Met 1	Glu	Pro	Lys	Ser 5	Ala .	As p	Lys		His 10	Thr	Cys	Pro	Pro	Cys 15	Pro	
Ala	Pro	Glu	Leu 20	Leu	GJA	Gly	Pro	Ser 25	Val	Phe	Leu	Phe	Pro 30	Pro	Lys	
Pro	Lys	Авр 35	Thr	Leu	Met	Ile	Ser 40	Arg	Thr	Pro	Glu	Val 45	Thr	Cys	Val	
Val	Val 50	Asp	Val	Ser	His	Glu 55	As p	Pro	Glu	Val	Lys 60	Phe	As n	Trp	Tyr	
Val 65	Asp	Gly	Val	Glu	Val 70	His	Asn	Ala	Lys	Thr 75	Lys	Pro	Arg	Glu	Gl u 80	
Gln	Tyr	Asn	Ser	Thr 85	Tyr	Arg	Val	Val	Ser 90	Val	Leu	Thr	Val	Leu 95	His	
Gln	Asp	Trp	Leu 100	Asn	Gly	Lys	Glu	Tyr 105		Сув	Lys	Val	Ser 110		Lys	

- 62 -

(2)	INFORMATION	FOR	SEC	 NO.	13.

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1135 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
 - (B) LOCATION: 4
 - (D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CATATGGACA AAACTCACAC ATGTCCACCT TGTCCAGCTC CGGAACTCCT GGGGGGTCCT 6.0 TCAGTCTTCC TCTTCCCCCC AAAACCCAAG GACACCCTCA TGATCTCCCG GACCCCTGAG 120 GTCACATGCG TGGTGGTGGA CGTGAGCCAC GAAGACCCTG AGGTCAAGTT CAACTGGTAC 180 GTGGACGGCG TGGAGGTGCA TAATGCCAAG ACAAGCCGC GGGAGGAGCA GTACAACAGC 240 ACGTACCGTG TGGTCAGCGT CCTCACCGTC CTGCACCAGG ACTGGCTGAA TGGCAAGGAG 300 TACAAGTGCA AGGTCTCCAA CAAAGCCCTC CCAGCCCCCA TCGAGAAAAC CATCTCCAAA 360 GCCAAAGGC AGCCCGAGA ACCACAGGTG TACACCCTGC CCCCATCCCG GGATGAGCTG 420 ACCAAGAACC AGGTCAGCCT GACCTGCCTG GTCAAAGGCT TCTATCCCAG CGACATCGCC 480 GTGGAGTGGG AGAGCAATGG GCAGCCGGAG AACAACTACA AGACCACGCC TCCCGTGCTG 540 GACTECGACG GETECTTETT CETETACAGE AAGCTCACCG TGGACAAGAG CAGGTGGCAG 600 CAGGGGAACG TOTTOTCATG CTCCGTGATG CATGAGGCTC TGCACAACCA CTACACGCAG 660 AAGAGCCTCT CCCTGTCTCC GGGTAAAGTA CCGATCCAGA AAGTTCAGGA CGACACCAAA 720 ACCITAATTA AAACGATCGT TACGCGTATC AACGACATCA GTCACACCCA GTCGGTGAGC 780 TCTAAACAGA AAGTTACAGG CCTGGACTTC ATCCCGGGTC TGCACCCGAT CCTGACCTTG 840 TCCAAAATGG ACCAGACCCT GGCTGTATAC CAGCAGATCT TAACCTCCAT GCCGTCCCGT 900 AACGTTATCC AGATCTCTAA CGACCTCGAG AACCTTCGCG ACCTGCTGCA CGTGCTGGCA 960 TTCTCCAAAT CCTGCCACCT GCCATGGGCT TCAGGTCTTG AGACTCTGGA CTCTCTGGGC 1020

GGGGTCCTGG AAGCATCCGG TTACAGCACC GAAGTTGTTG CTCTGTCCCG TCTGCAGGGT 1080

- 63 -

TCCCTTCAGG ACATGCTTTG GCAGCTGGAC CTGTCTCCGG GTTGTTAATG GATCC	1135
(2) INFORMATION FOR SEQ ID NO:14:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1135 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GTATACCTGT TTTGAGTGTG TACAGGTGGA ACAGGTCGAG GCCTTGAGGA CCCCCCAGGA	60
AGTCAGAAGG AGAAGGGGGG TTTTGGGTTC CTGTGGGAGT ACTAGAGGGC CTGGGGACTC	120
CAGTGTACGC ACCACCACCT GCACTCGGTG CTTCTGGGAC TCCAGTTCAA GTTGACCATG	180
CACCTGCCGC ACCTCCACGT ATTACGGTTC TGTTTCGGCG CCCTCCTCGT CATGTTGTCG	240
TGCATGGCAC ACCAGTCGCA GGAGTGGCAG GACGTGGTCC TGACCGACTT ACCGTTCCTC	300
ATGITCACGI TCCAGAGGIT GITTCGGGAG GGTCGGGGGT AGCTCTTTTG GTAGAGGTTT	360
CGGTTTCCCG TCGGGGCTCT TGGTGTCCAC ATGTGGGACG GGGGTAGGGC CCTACTCGAC	420

CTGAGGCTGC CGAGGAGAA GGAGATGTCG TTGAGTGGC ACCTGTTCTC GTCCACCGTC 600
GTCCCCTTGC AGAAGAGTAC GAGGCACTAC GTACTCCGAG ACGTGTTGGT GATGTGGGTC 660
TTCTCGGAGA GGGACAGAGG CCCATTCAT GGCTAGGTCT TTCAAGTCCT GCTGTGGTTT 720
TGGAATTAAT TTTGCTAGCA ATGCGCATAG TTGCTGTAGT CAGTGTGGGT CAGCCACTCG 780
AGATTTGTCT TTCAATGTCC GGACCTGAAG TAGGGCCCA ACGTGGGCTA GGACTGGAAC 840
AGGTTTTACC TGGTCTGGGA CCGCACATATG GTCGTCTAGA ATTGGAGGTA CGGCAGGGCA 900
TTGCAATAGG TCTAGAGATT GCTGGAGCTC TTGGAAGCCC TGGACGACCT GCACGACCCGT 960
AAGAGGTTTA GGACGGTGGA CGGTACCCGA AGTCCAGAAC TCTGAGGACCT GAGAGACCCG 1020

CCCCAGGACC TTCGTAGGCC AATGTCGTGG CTTCAACAAC GAGACAGGGC AGACGTCCCA 1080

TOGTTCTTGG TCCAGTCGGA CTGGACGGAC CAGTTTCCGA AGATAGGGTC GCTGTAGCGG

CACCTCACCC TCTCGTTACC CGTCGGCCTC TTGTTGATGT TCTGGTGCGG AGGGCACGAC

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540

PCT/US97/23183 WO 98/28427

- 64 -

AGGGAAGTCC TGTACGAAAC CGTCGACCTG GACAGAGGCC CAACAATTAC CTAGG 1135

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 374 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

- (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
- Met Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu
- Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
- Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser
- His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu 55
- Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr
- Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn
- Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro 100
- Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln 115
- Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val
- Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val 150

- 65 -

Glu	Trp	Glu	Ser	Asn 165	Gly	Gln	Pro	Glu	Asn 170	Asn	Tyr	Lys	Thr	Thr	Pro
Pro	Val	Leu	Asp 180	Ser	Asp	Gly	Ser	Phe 185	Phe	Leu	Tyr	Ser	Lys 190		Thr
Val	Asp	Lys 195	Ser	Arg	Trp	Gln	Gln 200	Gly	Asn	Val	Phe	Ser 205	СЛа	Ser	Val
Met	His 210	Glu	Ala	Leu	His	As n 215	His	Тут	Thr	Gln	Lys 220	Ser	Leu	Ser	Leu
Ser 225	Pro	Gly	Lys	Val	Pro 230	Ile	Gln	Lys	Val	Gln 235	Asp	Asp	Thr	Lys	Thr 240
Leu	Ile	Lys	Thr	11e 245	Val	Thr	Arg	Ile	Asn 250	Asp	Ile	Ser	His	Thr 255	Gln
Ser	Val	Ser	Ser 260	Lys	Gln	Lys	Val	Thr 265	Gly	Leu	As p	Phe	11e 270	Pro	Gly
Leu	His	Pro 275	Ile	Leu	Thr	Leu	Ser 280	Lys	Met	Asp	Gln	Thr 285	Leu	Ala	Val
Tyr	Gln 290	Gln	Ile	Leu	Thr	Ser 295	Met	Pro	Ser	Arg	As n 30 0	Val	Ile	Gln	Ile
Ser 305	Asn	Asp	Leu	Glu	Asn 310	Leu	λrg	λsp	Leu	Leu 315	His	Val	Leu	Ala	Phe 320
Ser	Lys	Ser	Cys	His 325	Leu	Pro	Trp	Ala	Ser 330	Gly	Leu	Glu	Thr	Leu 335	Asp
Ser	Leu	Gly	Gly 340	Val	Leu	Glu	Ala	Ser 345	Gly	Tyr	Ser	Thr	Glu 350	Val	Val
Ala	Leu	Ser 355	λrg	Leu	Gln	Gly	Ser 360	Leu	Gln	λsp	Net	Leu 365	Trp	Gln	Leu
λsp	Leu 370	Ser	Pro	Gly	Сув										

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1135 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

- 66 -

(ix) FEATURE:

(A) NAME/KEY: misc feature

(B) LOCATION: 4

(D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CATATGGACA AAACTCACAC ATGCCCACCG TGCCCAGCTC CGGAACTCGA AGGTGGTCCG 60 TOAGTOTTOC TOTTCCCCCC AAAACCCAAG GACACCCTCA TGATCTCCCG GACCCCTGAG 120 GTCACATGCG TGGTGGTGGA CGTGAGCCAC GAAGACCCTG AGGTCAAGTT CAACTGGTAC 180 GTGGACGGCG TGGAGGTGCA TAATGCCAAG ACAAAGCCGC GGGAGGAGCA GTACAACAGC 240 ACGTACCGTG TGGTCAGCGT CCTCACCGTC CTGCACCAGG ACTGGCTGAA TGGCAAAGCT 300 TACGCATGCG CGGTCTCCAA CAAAGCCCTC CCAGCCCCA TCGAGAAAAC CATCTCCAAA 360 GCCAAAGGC AGCCCGAGA ACCACAGGTG TACACCCTGC CCCCATCCCG GGATGAGCTG 420 ACCARGANC AGGTCAGCCT GACCTGCCTG GTCAAAGGCT TCTATCCCAG CGACATCGCC 480 GTGGAGTGGG AGAGCAATGG GCAGCCGGAG AACAACTACA AGACCACGCC TCCCGTGCTG 540 GACTCCGACG GCTCCTTCTT CCTCTACAGC AAGCTCACCG TGGACAAGAG CAGGTGGCAG 600 CAGGGGAACG TCTTCTCATG CTCCGTGATG CATGAGGCTC TGCACAACCA CTACACGCAG 660 AAGAGCCTCT CCCTGTCTCC GGGTAAAGTA CCGATCCAGA AAGTTCAGGA CGACACCAAA 720 ACCITAATTA AAACGATCGT TACGCGTATC AACGACATCA GTCACACCCA GTCGGTGAGC 780 TOTARACAGA AAGTTACAGG COTGGACTTC ATCCCGGGTC TGCACCCGAT COTGACCTTG 840 TCCAAAATGG ACCAGACCCT GGCTGTATAC CAGCAGATCT TAACCTCCAT GCCGTCCCGT 900 AACGTTATCC AGATCTCTAA CGACCTCGAG AACCTTCGCG ACCTGCTGCA CGTGCTGGCA 960 TICTCCAAAT CONCOACCT GCCATGGGCT TCAGGTCTTG AGACTCTGGA CTCTCTGGGC 1020 GGGGTCCTGG AAGCATCCGG TTACAGCACC GAAGTTGTTG CTCTGTCCCG TCTGCAGGGT 1080 TCCCTTCAGG ACATGCTTTG GCAGCTGGAC CTGTCTCCGG GTTGTTAATG GATCC 1135

- 67 -

	THEODINAMION	FOR	CEO	TD	NO.17
[2]	INFORMATION	FUR	SEU	110	NO:I/

(i) SPOUENCE CHARACTERISTICS:

(A) LENGTH: 1135 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GTATACCTGT TTTGAGTGTG TACGGGTGGC ACGGGTCGAG GCCTTGAGCT TCCACCAGGC 60 AGTCAGAAGG AGAAGGGGGG TTTTGGGTTC CTGTGGGAGT ACTAGAGGGC CTGGGGACTC 120 CAGTGTACGC ACCACCACCT GCACTCGGTG CTTCTGGGAC TCCAGTTCAA GTTGACCATG 180 CACCTGCCGC ACCTCCACGT ATTACGGTTC TGTTTCGGCG CCCTCCTCGT CATGTTGTCG 240 TOCATOGCAC ACCAGTOGCA GGAGTGGCAG GACGTGGTCC TGACCGACTT ACCGTTTCGA 300 ATGCGTACGC GCCAGAGGTT GTTTCGGGAG GGTCGGGGGT AGCTCTTTTG GTAGAGGTTT 360 CGGTTTCCCG TCGGGGCTCT TGGTGTCCAC ATGTGGGACG GGGGTAGGGC CCTACTCGAC 420 TGGTTCTTGG TCCAGTCGGA CTGGACGGAC CAGTTTCCGA AGATAGGGTC GCTGTAGCGG 480 CACCTCACCC TCTCGTTACC CGTCGGCCTC TTGTTGATGT TCTGGTGCGG AGGGCACGAC 540 CTGAGGCTGC CGAGGAAGAA GGAGATGTCG TTCGAGTGGC ACCTGTTCTC GTCCACCGTC 600 GTCCCCTTGC AGAAGAGTAC GAGGCACTAC GTACTCCGAG ACGTGTTGGT GATGTGCGTC 660 TTCTCGGAGA GGGACAGAGG CCCATTTCAT GGCTAGGTCT TTCAAGTCCT GCTGTGGTTT 720 TGGAATTAAT TTTGCTAGCA ATGCGCATAG TTGCTGTAGT CAGTGTGGGT CAGCCACTCG 780 AGATTTGTCT TTCAATGTCC GGACCTGAAG TAGGGCCCAG ACGTGGGCTA GGACTGGAAC 840 AGGTTTTACC TOGTCTGGGA CCGACATATG GTCGTCTAGA ATTGGAGGTA CGGCAGGGCA 900 TTGCAATAGG TCTAGAGATT GCTGGAGCTC TTGGAAGCGC TGGACGACGT GCACGACCGT 960 AAGAGGTTTA GGACGGTGGA CGGTACCCGA AGTCCAGAAC TCTGAGACCT GAGAGACCCG 1020 CCCCAGGACC TICGTAGGCC AATGTCGTGG CTTCAACAAC GAGACAGGGC AGACGTCCCA 1080 AGGGAAGTCC TGTACGAAAC CGTCGACCTG GACAGAGGCC CAACAATTAC CTAGG 1135

- 68 -

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 374 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein

(ix) FEATURE:

- (A) NAME/KEY: Protein
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Glu

1 10 15

Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys λsp Thr Leu 20 25 30

Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser 35 40 45

His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu 50 55 60

Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr 65 70 75 80

Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn

Gly Lys Ala Tyr Ala Cys Ala Val Ser Asn Lys Ala Leu Pro Ala Pro 100 105 110

Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln 115 120 125

Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val 130 135 140

Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val 145 150 150 155

Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro 165 170 175

- 69 -

Pro	Val	Leu	Asp 180	Ser	Asp	Gly	Ser	Phe 185	Phe	Leu	Tyr	Ser	Lys 190	Leu	Thr
Val	Asp	Lys 195	Ser	Arg	Trp	Gln	Gln 200	Gly	Asn	Val	Phe	Ser 205	Cys	Ser	Val
Met	His 210	Glu	Ala	Leu	His	Asn 215	His	Tyr	Thr	Gln	Lys 220	Ser	Leu	Ser	Leu
Ser 225	Pro	Gly	Lys	Val	Pro 230	Ile	Gln	Lys	Val	Gln 235	Asp	Asp	Thr	Lys	Thr 240
Leu	Ile	Lys	Thr	11e 245	Val	Thr	Arg	Ile	Asn 250	Asp	Ile	Ser	His	Thr 255	Gln
Ser	Val	Ser	Ser 260	Lys	Gln	Lys	Val	Thr 265	Gly	Leu	Asp	Phe	Ile 270	Pro	Gly
Leu	His	Pro 275	Ile	Leu	Thr	Leu	Ser 280	Lys	Met	Asp	Gln	Thr 285	Leu	Ala	Val
Tyr	Gln 290	Gln	Ile	Leu	Thr	Ser 295	Met	Pro	Ser	Arg	As n 300	Val	Ile	Gln	Ile
Ser 305	Asn	Asp	Leu	Glu	As n 310	Leu	Arg	Asp	Leu	Leu 315	His	Val	Leu	Ala	Phe 320
Ser	Lys	Ser	Cys	His 325	Leu	Pro	Trp	Ala	Ser 330	Gly	Leu	Gl u	Thr	Leu 335	λsp
Ser	Leu	Gly	Gly 340	Val	Leu	Glu	Ala	Ser 345	Gly	Tyr	Ser	Thr	Glu 350	Val	Val
Ala	Leu	Ser 355	Arg	Leu	Gln	Gly	Ser 360	Leu	Gln	Asp	Met	Leu 365	Trp	Gln	Leu
Asp	Leu 370	Ser	Pro	Gly	Cys										

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PCT/IIS97/23183 WO 98/28427

-70-

CLAIMS

A protein having a formula selected from the group consisting of: $R_1 - R_2$ and $R_1 - L - R_3$, wherein 5 R, is a Fc protein or analog thereof, R, is an OB protein or analog thereof, and L is a linker.

- The protein according to claim 1, where in the Fc, analog or derivative is selected from the group consisting of:
 - (a) the Fc amino acid sequences as set forth in SEO. ID. NOS.: 9, 12, 15 and 18:
 - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to SEO, ID, NO. 9):
 - one or more cysteine residues (i) replace by an alanine or serine residue:
 - one or more tyrosine residues (11) replaced by a phenylalanine residue;
 - (iii) the amino acid at position 5 replaced with an alanine:
 - the amino acid at position 20 (iv) replaced with glutamate;
- 25 121 the amino acid at position 103 replaced with an alanine;
 - the amino acid at position 105 (vi) replaced with an alanine;
 - (vii) the amino acid at position 107 replaced with an alanine;
 - (viii) the amino acids at positions 1, 2, 3, 4, or 5 deleted:
 - one or more residues replaced (ix) or deleted to ablate the Fc receptor binding site;

or deleted to ablate the complement (Clg) binding

one or more residues replaced

	site; and
	<pre>(xi) a combination of subparts i-x;</pre>
5	(c) the amino acid sequence of subparts (a)
	or (b) having a methionyl residue at the
	N-terminus;
	(d) the Fc protein, analog or derivative of
	any of subparts (a) through (c) comprised of a
10	chemical moiety connected to the protein moiety;
	(e) a derivative of subpart (d) wherein said
	chemical moiety is a water soluble polymer moiety;
	(f) a derivative of subpart (e) wherein said
	water soluble polymer moiety is polyethylene
15	glycol;
	(g) A derivative of subpart (e) wherein said
	water soluble polymer moiety is a polyamino acid
	<pre>moiety; and (h) a derivative of subpart (e) wherein said</pre>
20	water soluble polymer moiety is attached at solely
	the N-terminus of said protein moiety.
	the N cerumas of Sala protein morety.
	3. The protein according to claim 1, wherein
	the OB protein, analog or derivative is selected from
25	the group consisting of:
	(a) the amino acid sequence 1-146 as set
	forth in SEQ. ID. NO. 3 or SEQ. ID. NO. 6;
	(b) the amino acid sequence 1-146 as set
	forth in SEQ. ID. NO. 6 having a lysine residue at
30	position 35 and an isoleucine residue at position
	74;
	(c) the amino acid sequence of subpart (b)
	having a different amino acid substituted in one or
	more of the following positions (using the
35	numbering according to SEQ. ID. NO. 6): 4, 8, 32,

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33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145; (d) the amino acid sequence of subparts (a). (b) or (c) optionally lacking a glutaminyl residue at position 28: (e) the amino acid sequence of subparts (a). (b), (c), or (d) having a methionyl residue at the N-terminus. (f) a truncated OB protein analog selected from among: (using the numbering of SEO, ID, NO, 6 having a lysine residue at position 35, and an isoleucine residue at position 74): (i) amino acids 98-146 (ii) amino acids 1-32 (iii) amino acids 40-116 (iv) amino acids 1-99 and 112-146 (v) amino acids 1-99 and 112-146 having one or more of amino acids 100-111 sequentially placed between amino acids 99 and 112; and. (vi) the truncated OB analog of subpart (f) (i) having one or more of amino acids 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145 substituted with another amino acid: (vii) the truncated analog of subpart (f) (ii) having one or more of amino acids 4. 8 and 32 substituted with another amino acid: (viii) the truncated analog of subpart (f) (iii) having one or more of amino acids 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111 and 112 replaced with an ther amino acid;

(vix) the truncated analog of subpart
(f)(iv) having one or more of amino acids 4, 8, 32,

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- 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 112, 118, 136, 138, 142, and 145 replaced with another amino acid;
- (x) the truncated analog of subpart

 (f) (v) having one or more of amino acids 4, 8,32,

 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74,

 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111,

 112, 118, 136, 138, 142, and 145 replaced with another amino acid;
 - (xi) the truncated analog of any of subparts (f)(i)-(x) having an N-terminal methionyl residue;
 - (g) the OB protein or analog derivative of any of subparts (a) through (f) comprised of a chemical moiety connected to the protein moiety;
 - (h) a derivative of subpart (g) wherein said chemical moiety is a water soluble polymer moiety;
 - (i) a derivative of subpart (h) wherein said water soluble polymer moiety is polyethylene glycol;
 - (j) A derivative of subpart (h) wherein said water soluble polymer moiety is a polyamino acid moiety; and
- (k) a derivative of subpart (h) wherein said
 water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.
- 4. The protein of claim 1 wherein the linker sequence is one or more amino acids selected from the 30 group consisting of: Glycine, Asparagine, Serine, Threonine and Alanine.
 - 5. The protein of claim 1 wherein the linker is selected from the group consisting of:
- 35 (a) ala, ala, ala;
 - (b) ala, ala, ala, ala;

(c) ala, ala, ala, ala, ala; (6) glv. glv: (e) gly, gly, gly; gly, gly, gly, gly, gly; (f) 5 gly, gly, gly, gly, gly, gly, gly; (q) (h) gly-pro-gly; (i) gly, gly, pro, gly, gly; chemical moiety; and (i) (k) any combination of subparts (a) 10 through (j).

 A fusion protein comprising a Fc protein, analog or derivative thereof, fused to the N-terminus of an OB protein, analog or derivative thereof.

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- 7. A nucleic acid sequence encoding for a protein having the formula selected from the group consisting of: R₁ - R₂ and R₁ - L - R₂, wherein R₁ is a Fc protein or analog thereof, R₂ is an OB protein or analog thereof, and L is a linker.
- The nucleic acid sequence according to claim 7 encoding for a protein having a Fc, analog or
 derivative portion selected from the group consisting of:
 - (a) the Fc amino acid sequences as set forth in SEQ. ID. NOS.: 9, 12, 15 and 18;
 - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 9):
 - (i) one or more cysteine residues replace by an alanine or serine residue:

PCT/US97/23183

glycol;

moiety; and

	(ii) one or more tyrosine residues
	replaced by a phenylalanine residue;
	(iii) the amino acid at position 5
	replaced with an alanine;
5	(iv) the amino acid at position 20
	replaced with glutamate;
	(v) the amino acid at position 103
	replaced with an alanine;
	(vi) the amino acid at position 105
10	replaced with an alanine;
	(vii) the amino acid at position 107
	replaced with an alanine;
	(viii) the amino acids at positions 1,
	2, 3, 4, or 5 deleted;
15	(ix) one or more residues replaced
	or deleted to ablate the Fc receptor binding site;
	(x) one or more residues replaced
	or deleted to ablate the complement (C1q) binding
	site; and
20	<pre>(xi) a combination of subparts i-x;</pre>
	(c) the amino acid sequence of subparts (a)
	or (b) having a methionyl residue at the
	N-terminus;
	(d) the Fc protein, analog or derivative of
25	any of subparts (a) through (c) comprised of a
	chemical moiety connected to the protein moiety;
	(e) a derivative of subpart (d) wherein said
	chemical moiety is a water soluble polymer moiety;
	(f) a derivative of subpart (e) wherein said
30	water soluble polymer moiety is polyethylene

(g) A derivative of subpart (e) wherein said water soluble polymer moiety is a polyamino acid

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- (h) a derivative of subpart (e) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.
- 5 9. The nucleic acid sequence according to claim 7 encoding for a protein having an OB protein, analog or derivative portion selected from the group consisting of:
- (a) the amino acid sequence 1-146 as set 10 forth in SEQ. ID. NO. 3 or SEQ. ID. NO. 6;
 - (b) the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 6 having a lysine residue at position 35 and an isoleucine residue at position 74;
- (c) the amino acid sequence of subpart (b) having a different amino acid substituted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 6): 4, 8, 32,
 - 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145;
 - (d) the amino acid sequence of subparts (a),(b) or (c) optionally lacking a glutaminyl residue at position 28;
- (e) the amino acid sequence of subparts (a), (b), (c), or (d) having a methionyl residue at the N terminus.
 - (f) a truncated OB protein analog selected from among: (using the numbering of SEQ. ID. NO. 6 having a lysine residue at position 35, and an isoleucine residue at position 74):
 - (i) amino acids 98-146
 - (ii) amino acids 1-32
 - (iii) amino acids 40-116
- 35 (iv) amino acids 1-99 and 112-146

PCT/US97/23183

	(v) amino acids 1-99 and 112-146 having
	one or more of amino acids 100-111 sequentially
	placed between amino acids 99 and 112; and,
	(vi) the truncated OB analog of subpart
5	(f)(i) having one or more of amino acids 100, 102,
	105, 106, 107, 108, 111, 112, 118, 136, 138, 142,
	and 145 substituted with another amino acid;
	(vii) the truncated analog of subpart
	(f)(ii) having one or more of amino acids 4, 8 and
10	32 substituted with another amino acid;
	(viii) the truncated analog of subpart
	(f)(iii) having one or more of amino acids 50, 53,
	60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100,
	102, 105, 106, 107, 108, 111 and 112 replaced with
15	another amino acid;
	(vix) the truncated analog of subpart
	(f)(iv) having one or more of amino acids 4, 8, 32
	33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77
	78, 89, 97, 112, 118, 136, 138, 142, and 145
20	replaced with another amino acid;
	(x) the truncated analog of subpart
	(f)(v) having one or more of amino acids 4, 8,32,
	33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74,
	77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111
25	112, 118, 136, 138, 142, and 145 replaced with
	another amino acid;
	(xi) the truncated analog of any of
	subparts (f)(i)-(x) having an N-terminal methionyl
	residue;
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- (g) the OB protein or analog derivative of any of subparts (a) through (f) comprised of a chemical moiety connected to the protein moiety;
- (h) a derivative of subpart (g) wherein said chemical moiety is a water soluble polymer moiety;

PCT/US97/23183 WO 98/28427

-78-

(i) a derivative of subpart (h) wherein said water soluble polymer moiety is polyethylene glycol:

(j) A derivative of subpart (h) wherein said water soluble polymer moiety is a polyamino acid

(k) a derivative of subpart (h) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

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10. The nucleic acid sequence of claim 7 encoding for a protein with a linker sequence of one or more amino acids selected from the group consisting of: Gly, Asn, Ser, Thr and Ala.

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11. The nucleic acid sequence of claim 7 encoding for a protein with a linker selected from the group consisting of:

	(a)	ala, ala, ala;
20	(b)	ala, ala, ala, ala;
	(c)	ala, ala, ala, ala, ala;
	(D)	gly, gly;
	(e)	gly, gly, gly;
	(f)	gly, gly, gly, gly, gly;
25	(g)	gly, gly, gly, 'gly, gly, gly, gly;
	(h)	gly-pro-gly;
	(i)	gly, gly, pro, gly, gly;
	(E)	a chemical moiety; and
	(k)	any combination of subparts (a)

30 through (j).

12. A nucleic acid sequence encoding for a fusion protein having a Fc protein, analog or derivative thereof, fused to the N-terminus of an OB protein, 35 analog or derivative thereof.

-79-

13. A vector containing a nucleic acid sequence according to claims 7 or 12.

- 14. The vector of claim 13 wherein the vector is pAMG21 land the nucleic acid sequence according to claims 7 or 12.
 - 15. A prokaryotic or eukaryotic host cell containing the vector of claim 13.

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16. A process for producing a protein of claims 1 or 6 comprising the steps of culturing, under suitable conditions, the host cell of claim 15, and isolating the protein produced.

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- 17. The process of claim 16 further comprising the step of purifying the protein produced.
- 18. A pharmaceutical composition comprising 20 an effective amount of a protein according to claims 1 or 6, in a pharmaceutically acceptable diluent, adjuvant or carrier.
- 19. A method of treatment of a disorder
 25 selected from the group consisting of excess weight,
 diabetes, high blood lipid level, arterial sclerosis,
 arterial plaque, the reduction or prevention of gall
 stones formation, insufficient lean tissue mass,
 insufficient sensitivity to insulin, and stroke, wherein
 30 the method consists of administering a therapeutically
 effective amount of the protein according to claims 1 or
 6.

	CTCCCTGCCGTCCCAGAACGTTCTTCAGATCGCTAACGACCTCGAGAACCTTCGCGACCT	α C C
249	GAGGGCCCAGGGTCTTCCAAGAACTCTACCGATTGCTGGAGCTCTTGGAAGCGCTGGA S L P S Q N V L Q I A N D L E N L R D L	
309	GCTGCACCTGCTGGATTCTCCAAATCCTGCTCCCTGCCCCAACCTCAGGTCTTCAGAA 309 -++	368
369	ACCGGAATCCCTGGAGGTCCTGGAAGCATCCCTGTACAGCACCGAAGTTGTTGCTCT 369 +++	428
429	GTCCCGTCTGCAGGTTCCCTTCAGGACATCCTTCAGCAGCTGCACGCTTTCTCCGGAATG 429 -+	488
489	TTAATGGATCC 489 -+ AATTACCTAGG	

FIG 1E

FIG 2A

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420
         300
                                                                  360
                                                                    CTGGAGCTCTTGGAAGCGCTGGACGACGTGCACGACCGTAAGAGGTTTAGGACGGTGGAC
                                                                            GGTACCCGAAGTCCAGAACTCTGAGACCTGAGAGACCCGCCCCAGGACCTTCGTAGGCCA
                                                                                                                 TACAGCACCGAAGTIGITGCTCTGTCCCGTCTGCAGGGTTCCCTTCAGGACATGCTTTGG
                                                                                                                            ATGTCGTGGCTTCAACAACGAGACAGGCCAGACGTCCCAAGGGAAGTCCTGTACGAAACC
GACCTCGAGAACCTTCGCGACCTGCTGCTGCTGCTGGCATTCTCCAAATCCTGCCACCTG
                                                         CCATGGGCTTCAGGTCTTGAGACTCTGGACTCTGGGCGGGGTCCTGGAAGCATCCGGT
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FIG 2B

FIG 3A-1

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CATATGGAACCCAAATCTTGTGACAAAACTCACACATGCCCACGTGCCCAGCACCTGAA ++	M E P K S C D K T H T C P P C P A P E	CTCCTGGGGGGACCGTCGGTCTTCCTCTCTCCCCCAAACCCAAGGACACCCTCATGATC	L I G G P S V F L F P P K P K D T L M I -	TCCCGGACCCCTGAGGTCACATGGGTGGTGGAGCGTGAGCCACGAAGACCCTGAGGTC	SRTPEVTCVVVDVSHEDPEV -	AAGTICAACIGGIACGIGGACGGCGIGGAGGIGCATAATGCCAAGACAAAGCCGCGGGAG81	TWYVDGVEVHNAKTKPRE-
CCAAA	×	GACCG	<u>a</u>	CTGAC	ш	IGGTA(A Y
SAAC	ы	9999	v	ACCC TGGC	-	AAC	DI N
ATG(Σ	CTG	ü))))))	ĸ	FITC	TTCAAG K F
CAT		CTC	ū	TCC	S	AAC .	T. A
-		61		.21		.81	

B	/	9	1

;	GAGCAGTACAACAGCACGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGG	
7 6 7	CTCGTCATGTCGTGCATGGCACACCAGTCGCAGGAGTGGCAGGACGTGGTCCTGACC	
	EQYNSTYRVVSVLTVLHQDW -	
2	CTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAGGCCCTCCCAGCCCCCATCGAG	
301	GACTTACCGTTCCTCATGTTCACGTTCCAGAGGTTGTTTCGGGAGGGTCGGGGGTAGCTC	
	LNGKEYKCKVSNKALPAPIE -	
3	AAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGTGTACACCCTGCCCCCA	
351		
	KTISKAKGQPREPQVYTLPP	
:	TCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCTGCCTG	
431	431AGGCCCTACTCGACTCGTCCAGTCGGACGGACGGACGGAC	
	SRDELTKN Q V S L T C L V K G F Y -	
:	CCCAGGGACATCGCCGTGGAGTGGGAAGCAATGGGCAGCGGGAGAACAACTACAAGACC	
481	GGGTCGCTGTAGCGCCACCTCACCCTCTCGTTACCCGTCGGCCTCTTGTTGATGTTCTGG	

-1G 3A-2

FIG 3B-1

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                                                                                                                                                                        720
                                                                                                                                                                                                                                             AACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGTACCGATCCAGAAAGTT
                                                                                                                                                                                                                                CAGGACGACACCAAAACCTTAAATTAAAACGATCGTTACGCGTATCAACGACATCAGTCAC
                    ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTTCTTCCTTCTACAGCAAGCTCACGGTGGAC
                                          TGCGGAGGGCACGACCTGAGGCTGCCGAGGAAGAAGAGGAGATGTCGTTCGAGTGGCACCTG
                                                                                        AASAGCAGGTGGCAGCAGGGGAACGTCTŤCTCATGCTCCGTGATGCATGAGGCTCTGCAC
                                                                                                               TTCTCGTCCACCGTCGTCCCTTGCAGAAGAGTACGAGGCACTACGTACTCCGAGACGTG
                                                                                                                                                                                  TTGGTGATGTGCGTCTTCTCGGAGAGGGACAGAGGCCCATTTCATGGCTAGGTCTTTCAA
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AGCTC	TCGAG	S	TTGTC	AACAG		CGTA	GCAT	Z Z	GCAT	CGTA	Ā	99999	0000	დ დ
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GTCC	CAG	S	CCTC	GGAC	.J	2000	Ö	<u>α</u>	CGT	GCA GCA	>	CIC	GAG	Ø
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                  AGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACCGTCGACCTGGACAGAGGCCCAACA
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FIG 4A-1

CTCCTGGGGGGTCCTTCAGTCTTCCTTCCCCCCAAAACCCAAGGACACCCTCATGATC

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GAGGACCCCCCAGGAAGTCAGAAGGAGAAAGGGGGGTTTTGGGTTCCTGTGGGAGTACTAG

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09 CATATGGAACCAAAATCTGCTGACAAAACTCACATGTCCACCTTGTCCAGCTCCGGAA **GTATACCTTGGTTTTAGACGACTGTTTTGAGT**GTGTACAGGTGGAACAGGTCGAGGCCTT SADKTHTCPPCPA × а ы

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241	GAGCAGTACAACAGCACGTACCGTGTGGTCAGGGTCCTCACCGTCCTGGACTGG	300
	EQYNSTYRVVSVLTVLHQDW	1
301	CTGAATGGCAAGGAGTACAAGTGCAAGATCTCCAACAAAAAGCCTTCCCAGCCCCCATCGAG GACTTACCGTTCCTCATGTTCACGTTCCAGAGGTTGTTTCGGGAGGGTCGGGGGGTAGCTC	360
	LNGKEYKCKVSNKALPAPIE	
361	AAAACCATCTCCAAAGGCCAAAGGCAGCCCGAGAACCAGGGGTGTACACCTGCCCCA ++++++++++++-+-	420
	KTISKAKG QPREPQVYTLPP	1
421	TCCCGGGATGAGCTGACCAGGTCAGCTGACCTGACCTGA	480
	SRDELTKNQVSLTCLVKGFY	ı
481	CCCAGCGACATCGCCGTGGAGTGGGAGGCAATGGGCAGCCGGAGAACAACTACAAGACC	540
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FIG 4A-2

Y K T -	ACCGTGGAC + 600 TGGCACCTG	T V D -	GCTCTGCAC + 660 CGAGACGTG	А L Н -	AACCACTACAGGCAGAAGAGCCTCTCCCTGTCTCGGGTAAAGTACCGATCCAGAAAGTT	O K V -	CAGGACGACCAAAACCTTAAAACGATCGTTACGCGTATCAAGGACATCAGTCAC	TAGTCAGTG	
E N	CAGCAAGCTC +- STCGTTCGAG	FFLYSKL	SATGCATGAG +- CTACGTACTC	ы Н	AGTACCGATC +- TCATGGCTAG	K V P I Q	TATCAACGAC	ATAGTTGCTC	
G G	TTCCTCTA(F Y	ATGCTCCGT	c s	CCGGGTAA	r D	GTTACGCG	SCAATGCGC	
s N	SCTCCTTC	S	CTTCTCA + GAAGAGI	Er (N	CCTGTCT	r s	AACGATC	TTGCTAC	
E W E	TCCGACGC -+ AGGCTGCC	S D G	GGGAACGI -+	N N	AGCCTCTC -+	S I S	TTAATTA	AATTAATT	
> 4	SCTGGAC	I D	SCAGCAGO	N O O	SCAGAAG	N N	CAAAACC	TTTTGG	
SDIAV	ACCCTCCCGTGCTGGACTCCGACGCTCCTTCTTCCTCTACAGCTCACCGTGGAC TGCGGAGGGCACGACGACGACGACGAAGAAGAAGAAGATCGTTCGATCGGTCCTG	TPPVLDS	AAGAGCAGGTGGCAGCAGGAGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCAC	8 8	AACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGTACCGATCCAGAAAGTT 	H Y T	CAGGACGACCAAAAACCTTAAAAACGATCGTTACGCGTATCAACGACATCAGTCAC	GTCCTGCTGTGGTTTTGGAATTAATTTTGCTAGCAATGCGCATAGTTGCTGTAGTCAGTG	
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FIG 4B-2

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FIG 4C

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TCCCGICTGCAGGGTTCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGGTTGT
                    AGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACCGTCGACCTGGACAGAGGCCCAACA
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FIG 5A-1

ANTATGGACADAAA TOAGTCTTCTCT TCAGTCTTCCTCT AGTCAGAGGAGA S V F L F GTCACATGCGTGG CAGTGTACGCTGG V T C V V T C V V T C V V T C V V T C C CAGTGTACGCTGG	CATATGGACAAAACTCACACATGTCCACCTTGTCCAGCTCCGGAACTCCTGGGGGGTCCT	GTATACCTGTTTTGAGTGTGTACAGGTGGAACAGGTCGAGGCCTTGAGGACCCCCCAGGA	M D K T H T C P P C P A P E L L G G P -	TCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAG	AGTCAGAAGGAGAGAGAGGAGTTCTGTGGGAGTALTAGAGGGCTGGGGGAGTTCTGTGGGGAGTACTAGAGGGGGGGG	GTCACATGCGTGGTGGTGGACGTGGAGAGACCTGAGGTCAAGTTCAACTGGTAC 121++ 180 CAGTGTACGCACCACCACCAGCACTGGTTCTGGGACTCCAGTTCAAGTTGACCATG	V T C V V D V S H E D P E V К F N W Y -	GTGGACGCCGTGGAGGTGCATAATGCCAAGACAAGGCCGCGGGAGGAGAGAGA	CACCTGCCGCACCTCCACGTATTACGGTTCTGTTTTCGGCGCCCTCCTCGTCATGTTGTCG	
AND K M D K TOAGTCTTCTC TOAGTCTTCTC AGTCAGAAGGAG S V F L GTCACATGCGTG	ACTCAC	TGAGTG	H	TTCCCC	AAGGGG F P	GTGGTG CACCAC	۸ ۸	GAGGTG	CTCCAC	
M D TOAGTCTI S V F GTCACATG OTTOAGTCT CAGTGTAC V T C CAGTGTAC CACCTGCC	CAAA	GTTT	×	CCTC	GGAG	CGTG -+	>	CGTG	GCAC	
121 121 181	CATATGGA	STATACCT	Σ	rcagrerr	AGTCAGAA S V F	GTCACATG 	O T O	GTGGACGG	CACCTGCC	

720 009 099 GACTCCGACGCCTCCTTCTTCCTCTACAGCAGCTCACCGTGGACAAGAGCAGGTGGCAG CAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCAG GTCCCCTTGCAGAAGAGTACGAGGCACTACGTACTCCGAGACGTGTTGGTGATGTGCGTC AAGAGCCTCTCCCTGTCTCCGGGTAAAGTACCGATCCAGAAAGTTCAGGACGACAAAA **ACCTTAATTAAAACGATCGTTACGCGTATCA**ACGACATCAGTCACACCCCAGTCGGTGAGC **TIGAGGCTGCCGAGGAAGAAGAGATGTCG**TTCGAGTGGCACCTGTTCTCGTCCACCGTC **ITCTCGGAGAGGACAGAGGCCCATTTCATG**GCTAGGTCTTTCAAGTCCTGCTGTGGTTT **tggaattaattttgctagcaatgcgcatag**ttgctgtagtcagtgtggggtcagccactcg S > > α н S ۵ z 0 × H Ω > H × × ü > S × H æ ø × н K ш н Ω z Д I z z ഗ Σ > н ш > × >œ Д ᆸ S v α [I4 υ Д U S Ŋ <u>[14</u> z S Ŀ S Ø O ω z Ω ល ы 601

FIG 5B-1

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	K M D Q T L A V Y Q Q I L T S M P S	1
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961	TICTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACTCTGGGACTCTCTGGGCCAAAGGCCTCTCTGGGCCTAAAGGCGGTAAGGCCCGAAGTCCAGAACTCTGAGACCTGAGAAGCCCG	1020
	S K S C H L P W A S G L E T L D S L	ı
1021		1080
	CCCCAGGACCTICGIAGGCCAATGICGIGGCTICAACAACGACAAGACGAGACG	1
	FIG. 5B.2	

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FIG 5C

TCCCTTCAGGACATGCTTTGGCAGCTGGACTGTCTCCGGGTTGTTAATGGATCC

LODMLWQLDLSPGC

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	GTATACCTGTTTTGAGTGTGTACGGGTGGCAGGCGTCGAGGCCTTCAGCTTCCACCAGGC	MDKTHTCPPCPAPELEGGP	TCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAG 61+ 120 AGTCAGAAGGAGAAAGGGGGTTTTGGGTTCCTGTGGGAGTACTAGAGGGCCTGGGGACTC	SVFLFPPKFKDTLMISRTPB-	GTCACATGCGTGGTGGTGGAGCGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTAC 121	VICVVVDVSHEDPEVKFNWY-	GTGGACGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGGAGGAGGAGGAGGAGG	CACCTGCCGCACCTCCACGTATTACGGTTCTGTTTCGGCGCCCTCCTCGTCATGTTGTCG	· S N K O H H H H H A H A H A H A H A H A H A H
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FIG 6A-1

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ACGTACCGTGTGGTCGTCCTCACCGTCCTGCACTGACTGGATGGCAAAGCT	TGCATGGCACACGGTGCCAGGAGTGCCTGGTCCTGACCTGACCTTGCGTTCCACTTTCCACTTTCCACTTGCACGTGCGACGTGGCTGCTGGTTCCACTTTCCACTTTCCA	TACGCATGCGCGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAACCATCTCCAAA 301+++ 360 ATGCGTACGCGCCAGAGGTTGTTTCGGAGGTCGGGGGTAGCTCTTTTGGTAGAGGTTT	Y A C A V S N K A L P A P I E K T I S K -	GCCAAAGGGCAGCCCGGAGAACCACAGGTGTACACCCTGCCCCATCCCGGGATGAGCTG 361++++ 420 cggtttcccgtcggggctctttggtgtccAcatGTGGGACGGGAGTAGGGCCCTACTCGAC	AKGOPREPOVYTLPPSRDEL -	ACCAAGAACCAGGTCAGCCTGACCTGGTCAAAGGCTTCTATCCCAGCGACATGGC 421+ 480 TGGTTCTTGGTCGAGGACGGACGACGATTTCCGAAGATAGGGTCGCTGTAGCGG	KNQVSLTCLVKGFYPSDIA -
CCGTK	GGCA.	ATGC TACG	O	AGGG	O	GAAC	z
GGTA	3CATO	ACGC TGCG	Ø	CCAA GGTT	×	CCAA	Ŧ
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FIG 6A-2

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CACCTCACCCTCTCGTTACCCGTCGTCGTTGTTGATGTTCTGGTGCGGAGGGCACGAC

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FIG 6B-1

TCTAAACAGAAAGTTACAGGCCTGGACTTCATCCCGGGTCTGCACCCGATCCTGACCTTG	K Q K V T G L D F I P G L H P I L T L -	TCCAAAATGGACCAGACCTGGCTGTATACCAGCAGATCTTAACCTCCATGGCGTCGCGT AGGTTTTACCTGGTCTGGGACCGACATATGGTCGTTAGAAGTTAGGAGGTACGGCAGGGCA	K M D Q T L A V Y Q Q I L T S M P S R -	AACGTTATCCAGATCTCTAACGACCTCGAGAACCTTGGGGCCTGCTGCTGCTGCTGGCGCA 	VIQISNDLENLRDLLHVLA -	TTCTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACTCTGGACTCTGGGC+	S K S C H L P W A S G L E T L D S L G .	GGGGTCCTGGAAGCATCGGTTACAGCACGGAAGTTGTTGTTGTTGTGTCCGTCTGCAGGGT	V L E A S G Y S T E V A L S R L Q G
CCTAAA		CCCAAA.	×	AACGTT		TTCTCC 	FI S	SGGGTC	0 v
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FIG 6C

1081 TCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGTTAATGGATCC **AGGGAAGTCCTGTACGAAACCGTCGACCTGGACAGA**GGCCCAACAATTACCTAGG U Ġ а S ᆸ Ω ы Ø 3 ᆸ Σ Ω α ы S

INTERNATIONAL SEARCH REPORT

Inter anal Application No PCT/US 97/23183

A CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/62 C07K14/575 A61K38/17

According to International Patent Classification (IPC) or to both national classification and IPC

8. IPELDS SEARCHED

Maximum accommentation reserved (classification system followed by classification symbols)

IPC 6 C12N C07K A61K

Documentation searched other than mammum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

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Date of th	e actual completion of the international search	Date of mailing of the international se	serah report
	3 April 1998	3 0, 04, 98	
Neme an	i mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2	Authorized officer	
	NL - 2280 HV Rijendik Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Hermann, R	

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ategory *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
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Х	WO 97 00319 A (SMITHKLINE BEECHAM PLC) 3 January 1997	1-19
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